

## Research Article

# Isolation, Anti-Biotic Sensitivity and Biochemical Characterization of Fruit Spoiling Bacteria from Different Markets of Guwahati City

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### Abstract

There is potentiality for a wide range of vegetables and fruit products to become contaminated with microorganisms. The range of microorganisms associated with outbreaks linked to fresh product encompasses bacteria, viruses and parasites. This study was undertaken to investigate microbial safety aspects of some fruits such as Mango (*Mangifera indica*), Plum (*Prunus* sp.), Guava (*Psidium guajava*), Litchi (*Litchi chinensis*), Grapes (*Vitis vinifera*), Apple (*Malus domestica*) and Banana (*Musa acuminata*).

The presence of different bacteria which were responsible for spoiling fruits was confirmed by various biochemical and microbial tests. The microorganisms isolated were characterized by VITEK analysis and identified. The results showed the presence of the bacteria like *Enterobacter cloacae* complex, *Providencia rettgeri*, *Pantoea* spp and *Serratia plymuthica*. The isolated strains were then tested against 19 different antibiotics to look into their antibiotic susceptibility. The fruits in Guwahati region are mainly spoiled by *Enterobacter cloacae* complex, though other organisms are also there such as *Pantoea* spp, *Providencia rettgeri*, *Serratia plymuthica*. Methods to control these microbes need to be framed and applied for long term storage of the fruits.

**Keywords:** Antibiotic; Fruit Spoilage Microbes; Guwahati; Vitek

### Introduction

Fruits are vital sources of nutrients to human beings. They give the body the necessary vitamin and minerals in the right proportion for human growth and development. Fruits are seasonal and highly perishable in nature. The fruits that are cultivated in a natural environment are exposed to microbial contamination. They begin to deteriorate shortly after harvest. The fruits that are cultivated in the nature are more susceptible to the microbial attacks. Fruits may become contaminated with harmful pathogens and spoilage microorganisms either during their growing in fields, orchards, vineyards, or greenhouses, or during harvesting, postharvest handling, and distribution.

Sometimes spoilage of fruits also depends on microbial growth depends on various factors such as temperature, pH and

moisture of the particular region. Biochemical and microbial spoilage are main two causes for their spoilage. Due to the spoilage, the nutritional value and the market value of the fruits also gets deteriorated. It is estimated one-fourth of the harvested fruits are spoiled before consumption. Biochemical spoilage is caused by microorganisms like bacterial, yeast and moulds. Many microorganisms that are responsible for the spoilage of fruits are also harmful to human when consumed.

These may include respiratory failure, urinary infection, food poisoning, stomachache, fever, nausea etc. Over the years, there has been an increased demand to isolate and identify the microorganisms associated with the spoilage so that it can be controlled [1,2]. This study was undertaken to isolate and identify microorganisms that are associated or responsible for the spoilage of ripened fruits sold in Narengi and Paltan bazaar markets within Guwahati region. In addition, the study investigated the susceptibility of the microorganisms to some antibiotics. *Enterobacter* species are known to be associated with high resistance to most of the

antimicrobial agents [3]. Numerous studies on the mutation and its association with drug resistance have been reported [4-7]. Similar experience in terms of resistance was also experienced in the species isolated from food stuff [8-11]. The experiment was carried out to estimate and identify the fruit spoiling microbes in Guwahati.

## Materials and Methods

The experiments were conducted in the Department of Biotechnology, Assam Down Town University, Panikhaiti, Guwahati, Assam and apart with identification were carried out in Assam Down Town Hospital, India. The fruits selected for the study were: Apple (*Malus domestica*), Mango (*Mangifera indica*), Banana (*Musa acuminata*), Litchi (*Litchi chinensis*), Guava (*Psidium guajava*), Grapes (*Vitis vinifera*), Plum (*Prunus cerasifera*) which are most frequently consumed by the people of the region.

### Collection of Deteriorated Samples

All samples of fruits were collected from two markets of Guwahati region, Narengi and Paltan Bazaar. The fruits selected were slightly spoiled ones. The samples were taken to the laboratory, washed and drained of water. A sterile blade and forceps were used to cut small section of the tissue containing both the healthy and the rotten portion. The cut portions were pounded with pestle and mortar to make paste. The pastes were kept in sterile tubes and labeled for future use. A total of 9 representative samples were tested.

### Isolation of Bacteria

The pour-plate method according to Harigan and McCane (1990) [12] was adopted. Using standard Microbiological technique (serial dilution), a tenfold dilution of 1g of the sample was carried out in 9ml of sterile water (this was the aliquot). The exponential dilution was carried out to the fourth factor ( $10^{-4}$ ). 1ml of the fourth factor was aseptically transferred and plated in duplicate sets using sterile molten lukewarm nutrient agar which was amended with 1%. The plates were allowed to set and were incubated at 37°C for 24 hours. Individual colonies were chosen and the top of an individual colony each was touched lightly with a sterile loop or needle and streak on a fresh agar plate [13,14]. The various colonies observed in the plates were distinguished on the basis of their cultural characteristics such as colony size, shape, color, consistency and hemolytic characteristics. Bacterial growth was sub-cultured on NA and was preserved.

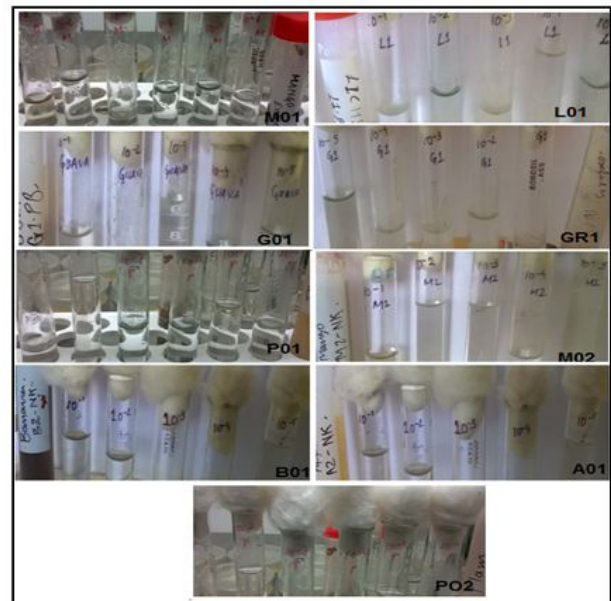
### Tests for Characterization of Bacterial Colonies

The tests to characterize the bacterial colonies were Gram staining, Catalase test, Methyl red test, Indole test, citrate utilization test, fermentation of carbohydrate test, triple sugar iron test, coagulase test, temperature assays, antibiotic tests and Vitek Test. The standard procedures for its test were performed [15-18]. As

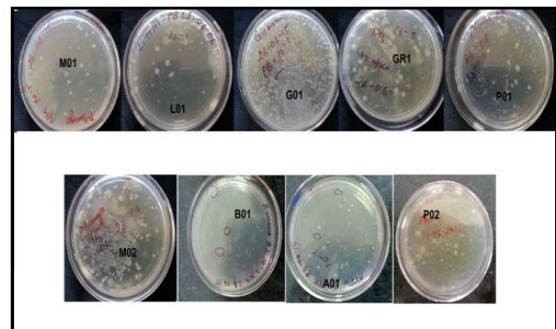
the current study was on representative samples hence statistical analysis was not undertaken.

## Results

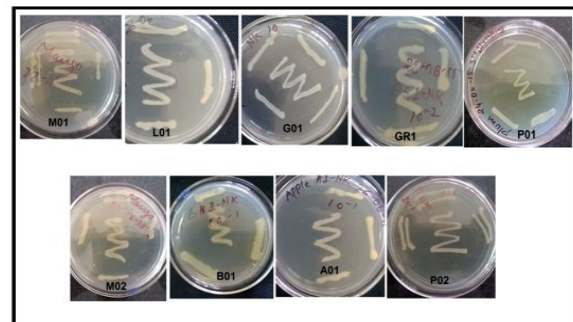
Collection of samples (spoiled fruits) and isolation of different bacteria from spoiled fruits could be seen from (Figure 1).



**Figure 1:** Serial Dilution Method.



**Figure 2:** Pour plate method.



**Figure 3:** Streaking Method.

The results for the biochemical tests are summarized in (Table 1). Most of the bacterial strains were positive for Catalase, citrate and coagulase tests [19]. The details of remaining tests have been shown in (Table 1).

Sl No.	Bacterial Strain	BIOCHEMICAL TEST RESULT							
		Catalase	METHYL RED TEST	INDOLE TEST	CITRATE UTILIZATION TEST	FERMENTATION OF CARBOHYDRATE	TRIPLE SUGAR IRON TEST		COAGULASE
							H <sub>2</sub> S	Gas Formation	
1.	M01	P	P	N	P	P	N	P	P
2.	L01	P	P	N	P	N	N	N	P
3.	G01	P	N	P	P	N	N	P	P
4.	GR1	P	P	N	P	N	N	P	P
5.	P01	P	P	N	P	N	N	N	P
6.	M02	P	N	P	P	N	N	P	P
7.	B01	P	P	N	P	P	N	P	P
8.	A01	P	P	P	P	P	N	P	P
9.	P02	P	P	N	P	P	N	P	P

Table 1: Biochemical Tests Results.

### Temperature Sensitivity Assay of Bacteria Strains

The results for temperature sensitivity shows response at 45°C for two out of the 10 studied strains of bacteria (Table 2 and Figure 4). The tests were found to be positive for all tested bacterial species at 30 and 40°C. From temperature sensitivity assay [21,22] it was found that all isolated bacterial strains grow optimally at 40°C.

Sl. No.	Bacterial strains	AREA	Temperature (°C)		
			30	40	45
1	M01	PALTAN BAZZAR	Y	Y	Y
2	L01		Y	Y	N
3	G01		Y	Y	N
4	GR1		Y	Y	N
5	P01		Y	Y	Y
6	M02	NARENGI	Y	Y	Y
7	B01		Y	Y	Y
8	A01		Y	Y	Y
9	P02		Y	Y	Y

Table 2: Temperature sensitivity assay of bacteria strains.

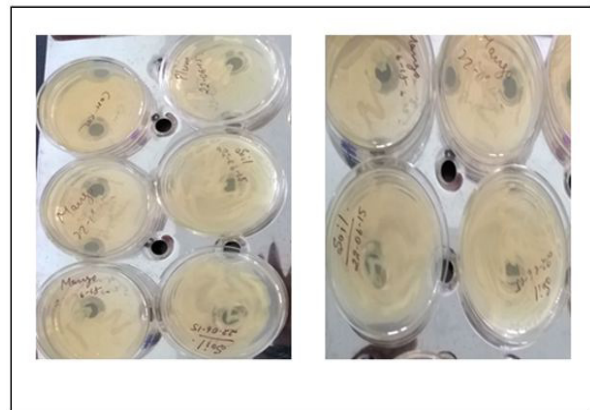


Figure 4: Temperature Sensitivity Assay.

### Identification of Isolated Bacterial Strains by Vitek Technique

In terms concentration, the highest number of bacterial load was observed of *Enterobacter* species (Table 3 and Figure 5). From the Vitek Test the species were identified as six species were identified as *Enterobacter cloacae* complex [23,25], one each of *Pantoea* spp, *Providencia rettgeri*, *Serratia plymuthica*.

Sl. NO	FRUITS	Bacterial Strain	Species name	Probability
1.	MANGO 1	M01	<i>Pantoea spp</i>	94%
2.	LITCHI	L01	<i>Providencia rettgeri</i>	99%
3.	GUAVA	G01	<i>Enterobacter cloacae complex</i>	97%
4.	GRAPES	GR1	<i>Enterobacter cloacae complex</i>	96%
5.	PLUM 1	P01	<i>Serratia plymuthica</i>	88%
6.	MANGO 2	M02	<i>Enterobacter cloacae complex</i>	99%
7.	BANANA	B01	<i>Enterobacter cloacae complex</i>	98%
8.	APPLE	A01	<i>Enterobacter cloacae complex</i>	99%
9.	PLUM 2	P02	<i>Enterobacter cloacae complex</i>	94%

Table 3: Result of Vitek Test.

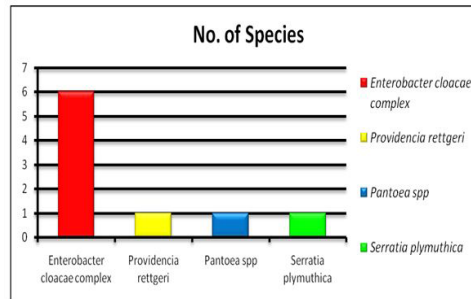


Figure 5: Graphical Representation of the No. of Species.

### Antibiotic Sensitivity Assay of Bacteria Strains

The results for antibiotic tests are summarized in Table 4. From the antibiotic sensitivity test we have tested 19 antibiotics on 9 strains of bacteria that were previously isolated and seen that most of the [26] strains are resistance to some of the antibiotics.

Sl. No.	SPECIMEN NAME	VITEK TEST RESULT	ANTIBIOTIC SENSITIVITY																		
			Alcipro 500	MonooF-O-200	Tremox (V-375) INCLAVE-375	Zefocep 200	Azomyein 500	Metron Fort	DOXY-100	Olox-200	L-Cin 500	Moxicap	OLOX-OZ	Cofmax-O	C'plex-TZ	NAR-TZ	Almox 500	Cofydoc-o 100 DT	Cephalex- 500	Ampicillin	Streptomycin
			A	M	T	Z	A1	A2	A3	A4	A5	A6	A7	A8	A9	A10	A11	A12	A13	A14	A15
1	M01	<i>Pantoea spp</i>	R	R	S	S	S	S	S	S	S	S	S	S	S	R	S	S	S	S	
2	L01	<i>Providencia rettgeri</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	
3	G01	<i>Enterobacter cloacae complex</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	
4	GR1	<i>Enterobacter cloacae complex</i>	S	R	S	S	R	S	S	S	S	S	S	S	S	S	S	S	S	S	
5	P01	<i>Serratia plymuthica</i>	S	S	S	S	S	-	S	S	S	S	S	S	S	S	S	S	S	S	
6	M02	<i>Enterobacter cloacae complex</i>	S	S	S	S	R	R	S	S	S	S	S	S	S	S	S	S	S	S	
7	B01	<i>Enterobacter cloacae complex</i>	S	R	S	S	S	S	S	S	S	R	S	S	S	S	S	S	S	S	
8	A01	<i>Enterobacter cloacae complex</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	
9	P02	<i>Enterobacter cloacae complex</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	

Table 4: Results for Antibiotic Tests.

### Conclusion

Fruits provide our body with essential nutrients; vitamins etc. However, there are some bacteria that cause spoilage of fruits. A conclusion achieved was that the fruits in Guwahati region are mainly spoiled by *Enterobacter cloacae complex*, though other organisms are also there such as *Pantoea spp*, *Providencia rettgeri*, *Serratia plymuthica* [27]. Again, presence of antibiotic resistant

strains needs to be dealt carefully. The study findings are of high importance as an appropriate method for controlling these micro organism needs to be adopted as the spoilage has economical im- plication.

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