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Review Article

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Genomic/Epigenomic Abnormality in Pediatric Cancer

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Abstract

Etiologies of pediatric cancers including childhood AML/ALL and brain tumors have been substantially explored. The findings have helped us illustrate a landscape of initiation and progression of tumor genesis, diagnosis/prognosis, and targeted treatments. Owing to the versatility of Next gen sequencing, unprecedented amount of data in cancer patients have been generated in order to characterize the complexity of genetic regulatory mechanisms. Many investigators have started to address cancerrelated questions using gene expression profiles, genetic variations, epigenetic profiles, and functional genetic regulatory elements at chromatin level. In this article, we systematically review how various genomic elements, such as gene fusion, mRNA and long non-coding RNA, may contribute to the development of pediatric cancers.

Conclusion: Full understanding of direct/indirect causative and consequent effects of aberrant patterns detected via largescale platforms in personalized medicine still remain elusive as pieces of puzzles. A better understanding of the role of these different genetic or genomic components may pave the way for identification of novel therapeutic targets.

Keywords: Lymphoblastic Leukemia Epigenetics; Mutation; Pathobiology; Pediatric

Cancer

Abbreviations:

AML Acute Myeloid Leukemia

AKML Acute Megakaryoblastic Leukemia

ALL Acute Lymphoblastic Leukemia

CNVs Copy Number Variations

CoBRA Combined Bisulfate Restriction Analysis

DIPG Diffuse Intrinsic Pontine Glioma

EWAS Epigenome-Wide Association Studies

GATK Genome-Analysis Toolkit

LncRNAs Long Non-Coding RNAs

LUNAR1 Leukemia-Induced Non-Coding

Activator RNA-1

Ph-like ALL Philadelphia Chromosome-Like Acute

SNVs Single Nucleotide Variations

SNPs Single Nucleotide Polymorphisms

T-ALL T cell Acute Lymphoblastic Leukemia

TSS Transcriptional Start Site

5mC 5-methylcytosine

5hmC 5-hydroxymethylcytosine

Background

Clinical heterogeneity has been thought to be a norm rather than an exception in many diseases -which indicates how different deregulated genetic pathways may lead to the same disease. Although many of previous studies[1-19] on the disease spectrum in pediatric AML/ALL and brain tumors have considerably focused on characterization of disease progression and targeted therapy, little has been known how abnormal patterns identified by super machinery Seq-based platforms explicitly work at cellular, molecular, organism al and functional level in

personalized medicine as the causal and consequent effects. A better understanding of each part of genetic or genomic susceptibility and their ensemble contributors such as gene fusions and disease specific epigenetic modification will enable to more precisely uncover the complexity of pediatric tumorigenesis and target novel treatments. Gene fusions results from the joining of parts of two previously separate genes. It commonly occurs when two different chromosomes break and parts fuse with one another to create a gene that is a hybrid and contains sequences from genes on the two different chromosomes [1,2,3,4,5]. The hybrid gene encodes a fusion protein that contains amino acid sequences derived from the parts of the two genes that were fused. Gene fusions resulting from chromosomal translocations are particularly common in cancers, especially hematological disorders and childhood sarcomas [1,3,6,7,8,9,10,11,12,13].

And expression of genes can be turned on or off or modified by factors other than an individual's DNA sequence. Although, theoretically, all cells in the same individual contain the same DNA sequences with the exception of mosaic phenomena, the pattern of expression of genes often vary by tissue because of epigenetic regulation [14].

The most common modification that influences expression of genes is DNA methylation. Genes with highly methylated promoters typically are not well expressed. Changes from normal patterns of DNA methylation of specific genes can also cause alterations in gene expression that are associated with disease. Recently it is also shown that DNA and RNA can be modified by methylation at 6' position N6-adenine methylation (N6-methyladenine; 6mA), which may harbor novel roles in gene regulation [5,18]. In addition to modification of nucleic acids, non-coding RNA also serves as another source of epigenetic mechanisms. Emerging evidence has suggested that non-coding RNA such as mRNA and long non-coding RNA (lncRNA) may play a key role in pediatric cancer [10,15,16,17,18,20].

In this article, we systematically review the significant interplay between genomic elements of interest and pediatric cancers that have not been thoroughly investigated thus far, such as gene fusions, epigenetic modifications including small and long non-coding RNAs.

Architecture of the Genome

Gene Fusion

Identification of the hybrid gene created by gene fusions as cause of a specific type of cancer can lead to the development of effective drugs. An example is the gene created by translocation between chromosomes 9 and 22 to create the "Philadelphia chromosome" characteristic of chronic myelogenous leukemia. The fusion gene (BCR/ABL) encodes an aberrant tyrosine kinase that causes leukemia. It has a catalytic site that differs from normal

enzyme. Specific inhibitors of the aberrant tyrosine kinase have been developed that do not inhibit the normal kinase and are effective personalized treatment for the disease [7]. The BCR/ABL fusion ontogeny has also been found in a small fraction of children with B-cell precursor acute lymphoblastic leukemia that has a poor prognosis [21].

The role of fusion genes in disease may be greatly underestimated because most gene expression profile studies have focused on normal versus diseased tissues. Searches for chromosomal translocations and indelsusing cytogenetic or special microarrays have not always been coupled with studies of gene expression. Recently RNA-seq has been adopted by investigators for transcript me studies followed by microarray analyses to search for chromosomal abnormalities. A study performed by Ataket al. [22]. identified SNVs underlying the over-expression of key driver genes of T cell acute lymphoblastic leukemia (T-ALL) using RNA-seq data. Masettiet al. [10] performed massively parallel sequencing of the transcript me of seven cases of cytogenetically normal pediatric AML and identified a novel recurrent CBFA2T3-GLIS2 fusion, which if present in leukemic cells of a patient predicted a poor outcome. They also identified a new fusion transcript, DHH-RHEBL1, in severalCBFA2T3-GLIS2-positive patients. Through the screening of a validation cohort of 55 pediatric AML patients, the authors identified a DHH-RHEBL1 fusion in 8 out of 20(40%) CBFA2T3-GLIS2- rearranged patients. The8 DHH-RHEBL1positive patients had a poorer survival than the 12 patients whose leukemic cells only harbored the CBFA2T3-GLIS2 rearrangement.

Epigenetic Modification of the Genome

DNA methylation

Studies of DNA methylation are often coupled with gene expression studies and genetic variation studies, as DNA methylation can regulate gene expression [23]. And SNPs also modify DNA methylation [24,25]. Methylation of DNA involves the conversion of cytosine to methyl cytosine (5mC) by adding a methyl group at 5' position at a site in DNA, and the oxidative intermediates generated during the de methylation processes(hydroxyl methyl, formyl, and carboxyl-cytosine) [26-28]. The cytosine nucleotide to be methylated is located next to a guanine nucleotide, i.e. in a CpGdinucleotide, although recent research has found methylated cytosine in other sequence contexts, such as CpA [26].

Epigenome-wide association studies (EWAS) hold promise for the detection of new regulatory mechanisms that may be susceptible to modification by environmental and lifestyle factors affecting disease [27]. Because of the rapid advances in sequencing technology, large numbers of methylated CpG sites can be identified across the entire genome. Studies of cancer frequently compare methylation sites in DNA from cancer tissues with sites in adjacent histologically tumor-free (i.e. normal) tissue. Both cancer

and normal tissues are typically available because of surgical resection of the cancer surrounded by normal tissue margins. DNA methylation profiles have also been used as molecular tools to subgroup cancer patients for personalized treatment [28-30]. In addition, utilizing cell-free DNA methylation as diagnostic tools and markers for treatment efficacy, especially for early detection of cancer, are being rapidly developed [31-33]. However, not many studies have focused on childhood leukemia and brain tumors.

It is important to note that current EWAS studies are limited by methods they used to measure DNA methylation (bead chip vs. sequencing-based methods, comparisons discussed in [34]. Most studies used arrays due to their cheaper cost, ease of use, consistency among large number of samples, and availability of well-established analytical tools. Even if Illumines bead arrays were utilized, the content of probes included (Golden gate vs. 27K vs. 450K) limited the number of CpG sites discovered. Furthermore, most of current studies replied on bisulfate treatment, which studied the sum of 5-methylcytosine (5mC), an oxidized form of 5-methylcytosine, and its base, 5hmC (5-hydroxymethylcytosine), instead of each modification individually.

Non-coding RNA

mRNAs may regulate gene expression by directly interfering transcription, or targeting post-translational processes including splicing, stability or translation of mRNA [35]. However, lncRNA is likely to regulate gene expression through modulating his tone modification [36], nucleosome positioning [37] and chromatin looping (enhancer RNA) [38-41]. As a result, they can target tumor-related signaling pathways, regulate inflammation and senescence-related genes [20], and promote proliferation and tumor growth by modulating cell cycle and inhibiting apoptosis [19, 42-43]. The roles of lncRNA in other aspects of cancer development have been extensively discussed [44]. Moreover, the use of mRNA and lncRNAas diagnostic and prognostic markers for pediatric tumors has been suggested [45,46].

In addition to pediatric brain tumors, emergent evidence also suggests the roles of mRNAs and long non-coding RNAs in T-ALL and AML. Trimarchiand colleagues (2014) [47]. Performed a relatively comprehensive characterization of long non-coding RNAs and proposed them as key multi-players leading to tumor genesis such as cellular homeostasis and transformation. In the study, a representative lncRNA referred to as LUNAR1(LeUkemia-induced Non-coding Activator RNA-1) has been thoroughly explored for its role in regulating neighboring genes such as IGF1R gene to promote Igf1 signaling and the growth of T-ALL by altering 3D chromatin structure and enhancer-specific features. Another study by Hirano et al. (2015) [48] has highlighted that lncRNA CCDC26 could be an effective therapeutic target to control myeloid leukemia cell growth via regulation of KIT expression, which is related to cell proliferation or survival. Furthermore, Emmrich

and colleagues [49] found that an interconnected functionality of lncRNAs MONC and MIR100HG plays a critical role as regulators of hematopoiesis and oncogenes, contributing to the maintenance of leukemic growth and therefore leading to the progressive Acute Megakaryoblastic Leukemia (AMKL). However, studies on mRNA and lncRNA at a genome-wide scale are still scarce and more comprehensive profiling is needed to further understand their roles in functional significance of cancer development, to evaluate their usage as diagnostic and prognostic markers, to develop more sophisticated and precise personalized treatments, and to determine subtype-specific features in tumor genesis [50,51]. In the following section, we will further discuss the role of DNA methylation in pediatric leukemia and brain tumor.

Disease-specific DNA methylation

The role of DNA methylation in cancers as well as diseases associated with imprinting has been extensively studied. With the emergence of high-throughput technologies, ranging from microarrays to next-gene sequencing, genome-wide scans to search for disease-related DNA methylation markers is now possible [52-58]. The interplay among DNA methylation sites, genetic variation, protein binding sites, and gene expression is a very active field of investigation.

Childhood leukemia

Nordlund et al identified genome-wide signatures associated with pediatric acute lymphoblastic leukemia (ALL) in 764 children at diagnosis and 27 children at relapse using the Infinium Human Methylation 450k Bead Chips [56]. In addition to 9,406 hyper ethylated CpG sites in ALL cells independent of cytogenetic background, each cytogenetic subtype displayed a unique set of hyper- and hypo methylated CpG sites. Further, these investigators identified DNA methylation signatures in cells at relapse that differed from those at the initial diagnosis from patients. Changes in DNA methylation signatures can predict disease relapse. Subtype-specific methylation patterns in promoters and enhancers were strongly correlated with gene expression. Hypo- and hyper-methylated sites are generally over-and under-expressed, respectively. Wong et al [58] studied DNA methylation profiles in archived bone marrow smears from 19 children with ETV6-RUNX1 (fusion gene that promotes survival of early lineage B-cells) positive pediatric pre-B cell ALL. Smears had been collected at diagnosis and remission and were used to derive a disease-specific DNA methylation profile. Gene signatures were confirmed in an independent cohort of 85 patients by quantitative analysis of DNA methylation and identification of differentially methylated CpG sites using SEQUENOM EpiTYPER. Methylation patterns at 15 loci were sufficient to discriminate leukemic from disease-free samples, regardless of the cytogenetic subtype of pre-B ALL. Aberrant methylation of TCF3, EGR4, BTG4 and hypomethylation of POU2AF1, TCF3, and RAG1 were found

in leukemic cells of ETV6-RUNX1 positive pediatric pre-B cell ALL cases. These genes are associated with B cell development. Chatterton et al [59] studied the DNA methylation and gene expression in 69 pediatric B-cell ALL patients (26 ETV6-RUNX1, 15 hyper diploids, and 28 "B-cell other") and 48 non-leukemic control samples. They identified 795 CpG sites with similarly deregulated DNA methylation levels common to three B-cell ALL subtypes, correlated with expression changes in 370 genes. In addition, there were also subtype-specific DNA methylation alterations with concurrent gene expression changes, which would help to increase understanding of the pathogenesis of each disease subtype. They can potential guide refinement of current treatment regimes tailored to each individual and can be used as robust biomarkers of disease progression states.

Figueroa et al. [60] studied the interaction among epigenetic markers, gene expression, and copy number variations in 137 B-lineage and 30 T-lineage childhoods lymphoblastic leukemia's using microarray-based technology. Consistent with other studies, this group found that different genetic subtypes of ALL were characterized by distinct DNA methylation signatures that exhibit significant correlation with gene expression profiles. There were epigenetic signatures common to all cases, with correlation to gene expression in 65% of these genes, suggesting that a core set of epigenetically deregulated genes is central to the initiation or maintenance of lymphoid transformation. Finally, genes with aberrant methylation were also targeted by recurring DNA copy number alterations in ALL, which suggests that expression of genes in CNVs are changed far more frequently than suggested by structural genomic analyses alone.

Childhood brain tumors

Although brain tumors are not as common as blood cancers in children, pediatric-onset brain tumors are characterized with a highest cancer-related mortality rate in children. The high mortality rate may be at least partly attributable to a limited number of clinical trials due to the scarcity of cases [61,62]. Furthermore, heterogeneous genetic mutations associated with some pediatriconset brain tumors may also partially explain why some single therapeutic agent cannot improve the clinical condition for a majority of cases. Omics-data facilitated by high-throughput sequencing have produced expansive evidence for novel epigenetic mechanisms underlying brain cancers, such as histone mutations, hijacking of enhancer elements, and novel oncogenic gene fusions - to name a few of them. For example, structural variants near the growth factor independent 1 family proto-oncogenes, GFI1 and GFI1B, may be active enhancer elements, including super-enhancers to cause medulloblastoma [59]. These findings exemplify how heterogonous the genetic factors associated with the susceptibility to pediatric-onset brain tumors could be.

The role of DNA methylation in childhood neuroblastoma has been explored by Lau and colleagues using customized Illumine Golden Gate methylation assays for 96 CpG sites within 48 candidate genes in primary neuroblastoma tumors from 131 children [60]. Levels of DNA methylation were validated in a subset of 48 patients using combined bisulfate restriction analysis (CoBRA) and bisulfate sequencing. The authors discovered that hyper methylation of FOLH1, MYOD1 and THBS1 are significant independent predictors of poorer clinical outcomes after adjusting for known prognostic factors. This study highlights the potential use of methylation profiling to identify prognostic markers and detect new therapeutic targets for selected subsets of patients. Studies of DNA methylation are often coupled with gene expression studies and genetic variation studies, as DNA methylation can regulate gene expression [63] and SNPs also modify DNA methylation [64-65]. Hovestadt et al. used Infinium Human Methylation 450k Bead Chips to study medulloblastoma and have successfully identified 48 CpG sites whose methylation can distinguish 4 subgroups of medulloblastoma [34]. Danielsson et al. further extended this study to all pediatric brain tumors and have developed a tool named MethPed that allow researchers and clinicians to classify patients into 9 brain tumor subtypes for further consideration of treatment development [37]. Fontebassoet al combined whole exome sequencing, gene expression and global DNA methylation profiles to study pediatric midline high-grade astrocytomas (including diffuse intrinsic pentane glioma or DIPG) [64]. Mutations in his tones have been found common to most tumors (37/40), which are correlated with common changes in DNA methylation. Recently, the deregulation of Loss of 5hmC in DIPG has been suggested [65] and genome-wide studies are hence warranted to examine the roles of 5mC and 5hmC in brain tumors.

Gene Expression Profiles

Comparing gene expression profiles of normal with diseased tissues has long been a common research method to study the pathophysiology of disease. Originally such studies were primarily conducted using expression microarrays. Use of microarrays is now typically coupled with studies using high throughput technologies such as RNA-seq[70-78]. These techniques have enabled the generation of lists of top putative candidates of differentially expressed genes between different groups in diseases of interest. Studies of gene expression generate information about alternative splicing of RNA transcripts, the role of non-coding genomic elements such as ncRNAs, ncDNAs, and microRNAs and epigenetic changes in the genome [52-54,66-72].

Leukemia's are the most common pediatric cancers. Several groups have studied gene expression profiles in B-cell and T-cell pediatric acute lymphoblastic leukemia at diagnosis and relapse. And researchers have conducted

integrative approaches between gene expression and epigenetic modifications of the genome [24,70,73-76]. Busche et al. [24] focused on pre-B ALL carrying the ETV6-RUNX1 fusion gene created by translocation between chromosomes 12 and 21. This study aims at profiling a meta-data between DNA methylation and transcript me. They demonstrated that genome-wide DNA methylation profiling could separate pre-B ALL into major subtypes with different responses to therapy and frequency of relapse. The quantification of methylation levels in CpG-islands around the transcriptional start site was statistically compared in Spearman correlation measurement with expression levels, resulting from the varying spectrum of correlations for each patient sample. Interestingly, they compared methylation levels with RNA-seq gene expression data in 17 patient samples and observed inverse correlation between methylation at CpG-sites located within the TSS1500 and its corresponding gene expression levels. They found the highest inverse correlations for CpG-sites located within exon1, and the next highest inverse correlations, in turn, at TSS (CpG sites within 1kb from transcriptional start site), and 5'-UTR, whereas it shows slightly significant positive correlations at 3'-UTR. Correlation analysis irrespective of subtype-specific molecular features demonstrated differential methylation of previously identified sub-type classifier for t (12; 21) such as BMP4, CELSR1, DSC3, and PON2 that are significantly correlated with transcript expression in ALL. Thus, they demonstrated that a combined strategy between methylome and transcript me analyses can identify potential drivers of leukemogenesis. Meyer et al. [77] generated transcript me profiles of matched diagnosis and relapse bone marrow specimens from ten individuals with pediatric B-ALL using RNA-seq and identified Single Nucleotide Variations (SNVs) and insertions/deletions (indels) by the means of Genomeanalysis Toolkit (GATK) and fusions by utilizing in-house pipeline, respectively. They identified 20 newly acquired non-synonymous mutations not present at initial diagnosis. At relapse two of the 10 patients had acquired new mutations in the 5'-nucleotidase NT5C2. Full-axon sequencing of NT5C2 was completed in 61 further relapse specimens, 5 of which had a new mutation in NT5C2. Collectively, the study showed that all individuals who harbored NT5C2 mutations relapsed early, indicating that mutations in NT5C2 are associated with the outgrowth of drug-resistant clones in ALL and a poor prognosis. Identification of epigenetic and genomic alterations that change expression profiles of genes in cancer cells from patients who have responded poorly to current therapies help identify targets for development of new drugs. More recently, Roberts et al. [78] explored Philadelphia chromosome-like acute lymphoblastic leukemia (Ph-like ALL) on the basis of Next gen transcript me, whole-genome, exome sequencing, microarray profiling, and functional and cytogenetic assays. In the study, they pointed out that kinase-activating alterations in more than 90% of patients with Ph-like ALL were observed with rearrangements and sequence mutations and the authors suggest that adding tyrosine

inhibitors to current therapy will be a better approach to increase the survival rate of this type of patients.

Genetic Regulatory Networks

Information on co-regulators and co-expression of genes in normal and disease states has increased interest in genetic regulatory networks. The generation of genetic variation/gene expression/DNA methylation profiles by either array or sequencing platforms provides the opportunity to define connectivity maps. This permits assignment of genetic variants, differentially expressed or methylated genes associated with pediatric disorders to specific biochemical pathways and regulatory networks [79-85]. These technologies have been applied in cancer and other diseases and have uncovered genetic regulatory mechanisms of action for targeting disease-specific genes. The complex underlying mechanisms and phenotypic outcomes for diseases such as cancer are derived through multi-layers of biological processes involved in development, cell-cycle, and epigenetic machinery with cis-regulating and trans-acting effects [4,24,26,52,53,55-60,67,68,70,71,73,76,79,80,82,83,86-99]. For example, large-scale sequencing of primary tumors from childhood medulloblastoma patients and their matched normal blood revealed that mutations in enzymes responsible for epigenetic and histone modifications are present in all subtypes of this disease, suggesting that these mutations are prime candidates of driver events [100-105]. This led to identification of genes and pathways that might be associated with poor outcomes and could be targets for new drugs to treat medullobastomas with specific genetic mutations. Transcriptional profiling of medulloblastoma cohorts identified several distinct molecular subgroups that vary with respect to their demographics, responses to therapy, and clinical outcomes [101,106-109]. Neuroblastoma, like medulloblastomas, is a cancer of nerve cells and has been studied in a similar fashion [110,110,111]. Molecular classification of medulloblastomas and neuroblastoma is possible and similar approaches have started to be applied to other pediatric diseases. Knowledge of genomic changes can be used to identify the biochemical pathways and regulatory networks that are affected and to identify potential targets for drug therapy [98,112]. Molecular classification of a patient's disease should improve choices of therapy at diagnosis, given a particular molecular subtype, and lead to development of new therapies for subtypes with a poor prognosis.

Concluding Remarks

The rapid advances of genomics genetics research have shed light on several novel therapeutic targets for pediatric cancers. In this systematic review, we have comprehensively reviewed various genomic elements associated with pediatric cancers. We have focused on ontogeny driver genes, mutations, epigenetic modifications regulating gene expression in protein-

coding genes and non-coding regions, aberrant patterns on gene expression profile data, post-transcriptional splicing procedures and chromosomal translocations of fusion genes. These genetic or genomic elements may serve as indicators to evaluate clinical stages and treatment options and responses. The accurate protocols for detection of diagnosis and prognosis biomarkers and treatments for specific cancers are still unclear and the well-defined subclassification of tumor types is generally lacking at present [113-115]. In addition, it still remains unclear how disease-associated variants, gene expression profiles, epigenetic patterns, and other genomic phenomena, influence each other in cells. It is still not clear to define explicitly what roles the abnormal patterns have in order to derive the complex multiple-steps in initiation and progression of tumors [23,94,100,102,103,116].

Above and beyond targeting genes and mutations at individual level, in the state-of-principle, further development of methods in system biological approaches to integrate single-layer analyses is crucial [8,23,79, 117-119]. More recently, a few studies have investigated pediatric cancers by integrating methylome and transcriptome analyses, combining analyses of whole genome-wide association studies/exome sequencing and expression profiles, exploring micro RNAs-RNAs interactions, and integrating transcriptome and proteomics studies. In order to decode the entire cascade of tumor genesis that is induced by accumulating effects from multi-layer events, it is not sufficient to study the intricate cancer biology with the regards to its functional roles by quantification at the single level alone.

There currently exists a significant gap to define the relationship between the changes of genomic abnormalities and their impact on tumor growth signaling pathways and responses to external stimuli such as targeted drugs. Hence, it is also very important to precisely account for appropriate experimental settings and validation steps, analytical and methodological pipelines, and well-balanced clinical applications in cohort studies with suitable sample sizes and proper selection of clinical informative measurements. Thus, above of all, we need efficient and systematic strategies to characterize deregulated physiological functions caused by somatic mutations, oncogenic fusions, and other abnormalities in genomic elements at sub-molecular, cellular, tissue-specific and organismal levels during the initiation and progression of pediatric tumors. A better understanding of the landscape of various genetic functional changes associated with pediatric on cogenesis will provide an initial key to personalized medicine.

References

 Aulmann S, Longerich T, Schirmacher P, Mechtersheimer G, Penzel R (2007) Detection of the ASPSCR1-TFE3 gene fusion in paraffinembedded alveolar soft part sarcomas. Histopathology 50: 881-886.

- Clark MJ, Homer N, O'Connor BD, Chen Z, Eskin A, et al. (2010) U87MG decoded: the genomic sequence of a cytogenetically aberrant human cancer cell line. PLoS Genet 6: 1000832.
- Francis RW, Thompson-Wicking K, Carter KW, Anderson D, Kees UR, et al. (2012) Fusion Finder: a software tool to identify expressed gene fusion candidates from RNA-Seq data. PLoS One 7: 39987.
- Liyanage VR, Jarmasz JS, Murugeshan N, Del Bigio MR, Rastegar M, et al. (2014) DNA modifications: function and applications in normal and disease States. Biology (Basel) 3: 670-723.
- Masetti R, Togni M, Astolfi A, Pigazzi M, Manara E, et al. (2013) DHH-RHEBL1 fusion transcript: a novel recurrent feature in the new landscape of pediatric CBFA2T3-GLIS2-positive acute myeloid leukemia. Oncotarget 4: 1712-1720.
- Hrzenjak A, Moinfar F, Tavassoli FA, Strohmeier B, Kremser ML, et al. (2005) JAZF1/JJAZ1 gene fusion in endometrial stromal sarcomas: molecular analysis by reverse transcriptase-polymerase chain reaction optimized for paraffin-embedded tissue. J Mol Diagn 7: 388-395.
- Jabbour E, Kantarjian H (2012) Chronic myeloid leukemia: 2012 update on diagnosis, monitoring, and management. Am J Hematol 87: 1037-1045.
- Ley TJ, Mardis ER, Ding L, Fulton B, McLellan MD, et al (2008) DNA sequencing of a cytogenetically normal acute myeloid leukemia genome. Nature 456: 66-72.
- Mardis ER, Ding L, Dooling DJ, Larson DE, McLellan MD, et al. (2009) Recurring mutations found by sequencing an acute myeloid leukemia genome. N Engl J Med 361: 1058-1066.
- Masetti R, Togni M, Astolfi A, Pigazzi M, Manara E, et al. (2013) DHH-RHEBL1 fusion transcript: a novel recurrent feature in the new landscape of pediatric CBFA2T3-GLIS2-positive acute myeloid leukemia. Oncotarget 4: 1712-1720.
- Shugay M, Ortiz de Mendibil I, Vizmanos JL, Novo FJ (2013) Oncofuse: a computational framework for the prediction of the oncogenic potential of gene fusions. Bioinformatics 29: 2539-2546.
- Togni M, Masetti R, Pigazzi M, Astolfi A, Zama D, et al. (2015) Identification of the NUP98-PHF23 fusion gene in pediatric cytogenetically normal acute myeloid leukemia by whole-transcriptome sequencing. J Hematol Oncol 8: 69.
- Zhou JB, Zhang T, Wang BF, Gao HZ, Xu X (2013) Identification of a novel gene fusion RNF213SLC26A11 in chronic myeloid leukemia by RNA-Seq. Mol Med Rep 7: 591-597.
- Liyanage VR, Jarmasz JS, Murugeshan N, Del Bigio MR, Rastegar M, et al. (2014) DNA modifications: function and applications in normal and disease States. Biology (Basel) 3: 670-723.
- Chakravadhanula M, Ozols VV, Hampton CN, Zhou L, Catchpoole D, et al. (2014) Expression of the HOX genes and HOTAIR in atypical teratoid rhabdoid tumors and other pediatric brain tumors. Cancer Genet 207:425-428.
- Jha P, Agrawal R, Pathak P, Kumar A, Purkait S et al. (2015) Genomewide small non coding RNA profiling of pediatric high-grade gliomas reveals deregulation of several mRNAs, identifies down regulation of snoRNA cluster HBII-52 and delineates H3F3A and TP53 mutant-specific miRNAs and snoRNAs. Int J Cancer 137: 2343-2353.

- Lee YY, Yang YP, Huang MC, Wang ML, Yen SH, et al. (2014) MicroR-NA142-3p promotes tumor-initiating and radioresistant properties in malignant pediatric brain tumors. Cell Transplant 23: 669-690.
- Miele E, Buttarelli FR, Arcella A, Begalli F, Garg N, et al. (2014) Highthroughput microRNA profiling of pediatric high-grade gliomas. Neuro Oncol 16: 228-240.
- Song H, Han LM, Gao Q, Sun Y (2016) Long non-coding RNA CRNDE promotes tumor growth in medulloblastoma. Eur Rev Med Pharmacol Sci 20: 2588-2597.
- Jones TA, Jeyapalan JN, Forshew T, Tatevossian RG, Lawson AR, et al. (2015) Molecular analysis of pediatric brain tumors identifies microRNAs in pilocytic astrocytomas that target the MAPK and NFkappaB pathways. Acta Neuropathol Commun 3: 86.
- van der Veer A, Zaliova M, Mottadelli F, De Lorenzo P, Te Kronnie G, et al. (2014) IKZF1 status as a prognostic feature in BCR-ABL1-positive childhood ALL. Blood 123: 1691-1698.
- Atak ZK, Gianfelici V, Hulselmans G, De Keersmaecker K, Devasia AG, et al. (2013) Comprehensive analysis of transcriptome variation uncovers known and novel driver events in T-cell acute lymphoblastic leukemia. PLoS Genet 9: 1003997.
- Huether R, Dong L, Chen X, Wu G, Parker M, et al. (2014) The landscape of somatic mutations in epigenetic regulators across 1,000 paediatric cancer genomes. Nat Commun 5: 3630.
- Busche S, Ge B, Vidal R, Spinella JF, Saillour V, et al. (2013) Integration of high-resolution methylome and transcriptome analyses to dissect epigenomic changes in childhood acute lymphoblastic leukemia. Cancer Res 73: 4323-4336.
- Zuckerman T, Rowe JM (2014) Pathogenesis and prognostication in acute lymphoblastic leukemia. F1000Prime Rep 6: 59.
- Patil V, Ward RL, Hesson LB (2014) The evidence for functional non-CpG methylation in mammalian cells. Epigenetics 9: 823-828.
- Nelson VR, Heaney JD, Tesar PJ, Davidson NO, Nadeau JH (2012)
 Transgenerational epigenetic effects of the Apobec1 cytidine deaminase deficiency on testicular germ cell tumor susceptibility and embryonic viability. Proc Natl Acad Sci USA 109: 2766-2773.
- Hovestadt V, Remke M, Kool M, Pietsch T, Northcott PA, et al. (2013) Robust molecular subgrouping and copy-number profiling of medulloblastoma from small amounts of archival tumour material using highdensity DNA methylation arrays. Acta Neuropathol 125:913-916.
- Nordlund J, Backlin CL, Zachariadis V, Cavelier L, Dahlberg J, et al. (2015) DNA methylation-based subtype prediction for pediatric acute lymphoblastic leukemia. Clin Epigenetics 7: 11.
- Danielsson A, Nemes S, Tisell M, Lannering B, Nordborg C, et al. (2015) MethPed: a DNA methylation classifier tool for the identification of pediatric brain tumor subtypes. Clin Epigenetics 7: 62.
- 31. Li Z, Guo X, Tang L, Peng L, Chen M, et al. (2016.) Methylation analysis of plasma cell-free DNA for breast cancer early detection using bisulfate next-generation sequencing. Tumour Biol.
- Qi Q, Pan YF, Shen JJ, Gu XQ, Han SW, et al. (2016) Circulating DNA for detection of gastric cancer. Eur Rev Med Pharmacol Sci 20: 2558-2564.

- Fernandes J, Michel V, Camorlinga-Ponce M, Gomez A, Maldonado C, et al. (2014) Circulating mitochondrial DNA level, a noninvasive biomarker for the early detection of gastric cancer. Cancer Epidemiol Biomarkers Prev 23: 2430-2438.
- Stirzaker C, Taberlay PC, Statham AL, Clark SJ (2014) Mining cancer methylomes: prospects and challenges. Trends Genet 30: 75-84.
- Abdel-Hameed EA, Ji H, Shata MT (2016) HIV-Induced Epigenetic Alterations in Host Cells. Adv Exp Med Biol 879:27-38.
- 36. Hajjari M, Salavaty A (2015) HOTAIR: an oncogenic long non-coding RNA in different cancers. Cancer Biol Med 8: 170.
- Prensner JR, Iyer MK, Sahu A, Asangani IA, Cao Q, et al. (2013) The long noncoding RNA SChLAP1 promotes aggressive prostate cancer and antagonizes the SWI/SNF complex. Nat Genet 45: 1392-1398.
- Orom UA, Derrien T, Beringer M, Gumireddy K, Gardini A, et al. (2010) Long noncoding RNAs with enhancer-like function in human cells. Cell 143: 46-58.
- Melo CA, Drost J, Wijchers PJ, van de Werken H, de Wit E, et al. (2013) eRNAs are required for p53-dependent enhancer activity and gene transcription. Mol Cell 49:524-535.
- Li W, Notani D, Ma Q, Tanasa B, Nunez E, et al. (2013) Functional roles of enhancer RNAs for oestrogen-dependent transcriptional activation. Nature 498: 516-520.
- Lai F, Orom UA, Cesaroni M, Beringer M, Taatjes DJ (2013) Activating RNAs associate with Mediator to enhance chromatin architecture and transcription. Nature 494: 497-501.
- Kaid C, Silva PB, Cortez BA, Rodini CO, Semedo-Kuriki P (2015) miR-367 promotes proliferation and stem-like traits in medulloblastoma cells. Cancer Sci 106: 1188-1195.
- 43. Panwalkar P, Moiyadi A, Goel A, Shetty P, Goel N, et al. (2015) MiR-206, a Cerebellum Enriched miRNA Is Downregulated in All Medullo-blastoma Subgroups and Its Overexpression Is Necessary for Growth Inhibition of Medulloblastoma Cells. J Mol Neurosci 56: 673-680.
- Evans JR, Feng FY, Chinnaiyan AM, (2016) The bright side of dark matter: IncRNAs in cancer. J Clin Invest 126: 2775-2782.
- Li Y, Jiang T, Shao L, Liu Y, Zheng C (2016) Mir-449a, a potential diagnostic biomarker for WNT group of medulloblastoma. J Neurooncol.
- 46. Serghiou S, Kyriakopoulou A, Ioannidis JP (2016) Long noncoding RNAs as novel predictors of survival in human cancer: a systematic review and meta-analysis. Mol Cancer 15: 50.
- Trimarchi T, Bilal E, Ntziachristos P, Fabbri G, Dalla-Favera R, et al. (2014) Genome-wide mapping and characterization of Notch-regulated long noncoding RNAs in acute leukemia. Cell 158: 593-606.
- 48. Hirano T, Yoshikawa R, Harada H, Harada Y, Ishida A, et al. (2015) Long noncoding RNA, CCDC26, controls myeloid leukemia cell growth through regulation of KIT expression. Mol Cancer 14: 90.
- Emmrich S, Streltsov A, Schmidt F, Thangapandi VR, Reinhardt D, et al. (2014) LincRNAs MONC and MIR100HG act as oncogenes in acute megakaryoblastic leukemia. Mol Cancer 13: 171.
- Mitra SA, Mitra AP, Triche TJ, (2012) A central role for long non-coding RNA in cancer. Front Genet 3:17.

- 51. Frediani JN, Fabbri M (2016) Essential role of miRNAs in orchestrating the biology of the tumor microenvironment. Mol Cancer 15: 42.
- Jones PA (2012) Functions of DNA methylation: islands, start sites, gene bodies and beyond. Nat Rev Genet 13: 484-492.
- Lau DT, Hesson LB, Norris MD, Marshall GM, Haber M, et al. (2012) Prognostic significance of promoter DNA methylation in patients with childhood neuroblastoma. Clin Cancer Res 18: 5690-5700.
- Nordlund J, Backlin CL, Wahlberg P, Busche S, Berglund EC, et al. (2013) Genome-wide signatures of differential DNA methylation in pediatric acute lymphoblastic leukemia. Genome Biol 14: 105.
- 55. Ueno H, Okita H, Akimoto S, Kobayashi K, Nakabayashi K, et al, (2013) DNA methylation profile distinguishes clear cell sarcoma of the kidney from other pediatric renal tumors. PLoS One 8: 62233.
- van Eijk KR, de Jong S, Boks MP, Langeveld T, Colas F, et al. (2012) Genetic analysis of DNA methylation and gene expression levels in whole blood of healthy human subjects. BMC Genomics 13:636.
- Wagner JR, Busche S, Ge B, Kwan T, Pastinen T, et al (2014) The relationship between DNA methylation, genetic and expression interindividual variation in untransformed human fibroblasts. Genome Biol 15: 37.
- Wong NC, Ashley D, Chatterton Z, Parkinson-Bates M, Ng HK, et al. (2012) A distinct DNA methylation signature defines pediatric pre-B cell acute lymphoblastic leukemia. Epigenetics 7:535-541.
- Chatterton Z, Morenos L, Mechinaud F, Ashley DM, Craig JM, et al. (2014) Epigenetic deregulation in pediatric acute lymphoblastic leukemia. Epigenetics 9: 459-467.
- Figueroa ME, Chen SC, Andersson AK, Phillips LA, Li Y, et al. (2013) Integrated genetic and epigenetic analysis of childhood acute lymphoblastic leukemia. J Clin Invest 123: 3099-3111.
- Northcott PA, Pfister SM, Jones DT (2015) Next-generation (epi) genetic drivers of childhood brain tumours and the outlook for targeted therapies. Lancet Oncol 16: 293-302.
- International Cancer Genome C, Hudson TJ, Anderson W, Artez A, Barker AD, et al. (2010) International network of cancer genome projects. Nature 464: 993-998.
- Northcott PA, Lee C, Zichner T, Stutz AM, Erkek S, et al (2014) Enhancer hijacking activates GFI1 family oncogenes in medulloblastoma. Nature 511: 428-434.
- 64. Fontebasso AM, Papillon-Cavanagh S, Schwartzentruber J, Nikbakht H, Gerges N, et al (2014) Recurrent somatic mutations in ACVR1 in pediatric midline high-grade astrocytoma. Nat Genet 46: 462-466.
- Johnson KC, Houseman EA, King JE, von Herrmann KM, Fadul CE, et al. (2016) 5-Hydroxymethylcytosine localizes to enhancer elements and is associated with survival in glioblastoma patients. Nat Commun 7: 13177.
- Fan M, Krutilina R, Sun J, Sethuraman A, Yang CH, et al. (2013) Comprehensive analysis of microRNA (miRNA) targets in breast cancer cells. J Biol Chem 288: 27480-27493.
- Grech G, Pollacco J, Portelli M, Sacco K, Baldacchino S, et al. (2014) Expression of different functional isoforms in haematopoiesis. Int J Hematol 99: 4-11.

- Gutschner T, Diederichs S (2012) The hallmarks of cancer: a long noncoding RNA point of view. RNA Biol 9:703-719.
- 69. Liu Q, Chen C, Shen E, Zhao F, Sun Z, et al. (2012) Detection, annotation and visualization of alternative splicing from RNA-Seq data with SplicingViewer. Genomics 99: 178-182.
- Stunnenberg HG, Hubner NC (2014) Genomics meets proteomics: identifying the culprits in disease. Hum Genet 133: 689-700.
- Sultan M, Schulz MH, Richard H, Magen A, Klingenhoff A, et al (2008) A global view of gene activity and alternative splicing by deep sequencing of the human transcriptome. Science 321: 956-960.
- 72. Burke MJ, Bhatla T (2014) Epigenetic modifications in pediatric acute lymphoblastic leukemia.Front Pediatr 2: 42.
- Girardot M, Cavaille J, Feil R (2012) Small regulatory RNAs controlled by genomic imprinting and their contribution to human disease. Epigenetics 7: 1341-1348.
- 74. Mullighan CG (2013) Genome sequencing of lymphoid malignancies. Blood 122: 3899-3907.
- Van der Meulen J, Van Roy N, Van Vlierberghe P, Speleman F (2014)
 The epigenetic landscape of T-cell acute lymphoblastic leukemia. Int J Biochem Cell Biol 53: 547-557.
- Meyer JA, Wang J, Hogan LE, Yang JJ, Dandekar S, et al (2013) Relapse-specific mutations in NT5C2 in childhood acute lymphoblastic leukemia. Nat Genet 45: 290-294.
- Roberts KG, Li Y, Payne-Turner D, Harvey RC, Yang YL, et al. (2014)
 Targetable kinase-activating lesions in Ph-like acute lymphoblastic leukemia. N Engl J Med 371: 1005-1015.
- Basso K, Margolin AA, Stolovitzky G, Klein U, at al. (2005) Dalla-Favera R, Reverse engineering of regulatory networks in human B cells. Nat Genet 37: 382-390.
- Belcastro V, Siciliano V, Gregoretti F, Mithbaokar P, Dharmalingam G, et al. (2011) Transcriptional gene network inference from a massive dataset elucidates transcriptome organization and gene function. Nucleic Acids Res 39: 8677-8688.
- 80. Blake JA, Ziman MR (2014) Pax genes: regulators of lineage specification and progenitor cell maintenance. Development 141: 737-751.
- Civelek M, Lusis AJ (2014) Systems genetics approaches to understand complex traits. Nat Rev Genet 15: 34-48.
- 82. Kavanagh T, Mills JD, Kim WS, Halliday GM, Janitz M (2013) Pathway analysis of the human brain transcriptome in disease. J Mol Neurosci 51: 28-36.
- 83. Rodriguez-Zas SL, Ko Y, Adams HA, Southey BR (2008) Advancing the understanding of the embryo transcriptome co-regulation using meta-, functional, and gene network analysis tools. Reproduction 135: 213-224
- Massague J (2008) A very private TGF-beta receptor embrace. Mol Cell 29: 149-150.
- 85. Wang Z, Lo HS, Yang H, Gere S, Hu Y, et al. (2003) Computational analysis and experimental validation of tumor-associated alternative RNA splicing in human cancer. Cancer Res 63: 655-657.

- Whichard ZL, Sarkar CA, Kimmel M, Corey SJ (2010) Hematopoiesis and its disorders: a systems biology approach. Blood 115: 2339-2347.
- 87. Almamun M, Levinson BT, van Swaay AC, Johnson NT, McKay SD, et al. (2015) Integrated methylome and transcriptome analysis reveals novel regulatory elements in pediatric acute lymphoblastic leukemia. Epigenetics 10: 882-890.
- Brait M, Sidransky D (2011) Cancer epigenetics: above and beyond. Toxicol Mech Methods 21: 275-288.
- 89. Chen RL, Guo W, Shi Y, Wu H, Wang J, et al. (2013) Computational identification of specific splicing regulatory elements from RNA-seq in lung cancer. Eur Rev Med Pharmacol Sci 17: 1716-1721.
- Chen S, Bellew C, Yao X, Stefkova J, Dipp S, et al. (2011) Histone deacetylase (HDAC) activity is critical for embryonic kidney gene expression, growth, and differentiation. J Biol Chem 286: 32775-32789.
- 91. Chen X, Hu Z, Wang W, Ba Y, Ma L, et al. (2012) Identification of ten serum microRNAs from a genome-wide serum microRNA expression profile as novel noninvasive biomarkers for nonsmall cell lung cancer diagnosis. Int J Cancer 130: 1620-1628.
- Cheng J, Guo JM, Xiao BX, Miao Y, Jiang Z, et al. (2011) piRNA, the new non-coding RNA, is aberrantly expressed in human cancer cells. Clin Chim Acta 412: 1621-1625.
- Ding L, Getz G, Wheeler DA, Mardis ER, McLellan MD, et al. (2008) Somatic mutations affect key pathways in lung adenocarcinoma. Nature 455:1069-1075.
- Graveley BR (2001) Alternative splicing: increasing diversity in the proteomic world. Trends Genet 17: 100-107.
- Hubertus J, Zitzmann F, Trippel F, Muller-Hocker J, Stehr M, et al. (2013) Selective methylation of CpGs at regulatory binding sites controls NNAT expression in Wilms tumors. PLoS One 8: 67605.
- 96. Lee YH, Kim JH, Song GG (2014) Genome-wide pathway analysis in neuroblastoma. Tumour Biol 35: 3471-3485.
- Liu H, D'Andrade P, Fulmer-Smentek S, Lorenzi P, Kohn KW, et al. (2010) mRNA and microRNA expression profiles of the NCI-60 integrated with drug activities. Mol Cancer Ther 9: 1080-1091.
- Ohshima J, Haruta M, Fujiwara Y, Watanabe N, Arai Y, et al. (2012) Methylation of the RASSF1A promoter is predictive of poor outcome among patients with Wilms tumor. Pediatr Blood Cancer 59: 499-505.
- Forbes SA, Bindal N, Bamford S, Cole C, Kok CY, et al. (2011) COS-MIC: mining complete cancer genomes in the Catalogue of Somatic Mutations in Cancer. Nucleic Acids Res 39: 945-950.
- Northcott PA, Korshunov A, Witt H, Hielscher T, Eberhart CG, et al. (2011) Medulloblastoma comprises four distinct molecular variants. J Clin Oncol 29: 1408-1414.
- 101. Pleasance ED, Cheetham RK, Stephens PJ, McBride DJ, Humphray SJ, et al. (2010) A comprehensive catalogue of somatic mutations from a human cancer genome. Nature 463: 191-196.
- 102. Pugh TJ, Weeraratne SD, Archer TC, Pomeranz Krummel DA, Auclair D, et al. (2012) Medulloblastoma exome sequencing uncovers subtype-specific somatic mutations. Nature 488: 106-110.

- 103. Robinson G, Parker M, Kranenburg TA, Lu C, Chen X, et al. (2012) Novel mutations target distinct subgroups of medulloblastoma. Nature 488: 43-48.
- 104. Thompson MC, Fuller C, Hogg TL, Dalton J, Finkelstein D, et al. (2006) Genomics identifies medulloblastoma subgroups that are enriched for specific genetic alterations. J Clin Oncol 24: 1924-1931.
- 105. Cho SG, Lee JW, Heo JS, Kim SY (2014) Gene Expression Change in Human Dental Pulp Cells Exposed to a Low-Level Toxic Concentration of Triethylene Glycol Dimethacrylate: An RNA-seq Analysis. Basic Clin Pharmacol Toxicol.
- 106. Northcott PA, Hielscher T, Dubuc A, Mack S, Shih D, et al. (2011) Pediatric and adult sonic hedgehog medulloblastomas are clinically and molecularly distinct. Acta Neuropathol 122: 231-240.
- 107. Tamayo P, Cho YJ, Tsherniak A, Greulich H, Ambrogio L, et al. (2011) Predicting relapses in patients with medulloblastoma by integrating evidence from clinical and genomic features. J Clin Oncol 29: 1415-1423.
- 108. Taylor P, Fangusaro J, Rajaram V, Goldman S, Helenowski IB, et al. (2012) REST is a novel prognostic factor and therapeutic target for medulloblastoma. Mol Cancer Ther 11: 1713-1723.
- 109. Das S, Bryan K, Buckley PG, Piskareva O, Bray IM, et al. (2013) Modulation of neuroblastoma disease pathogenesis by an extensive network of epigenetically regulated microRNAs. Oncogene 32: 2927-2936.
- Jones DT, Jager N, Kool M, Zichner T, Hutter B, et al. (2012) Dissecting the genomic complexity underlying medulloblastoma. Nature 488: 100-105.
- Hanash SM, Madoz-Gurpide J, Misek DE (2002) Identification of novel targets for cancer therapy using expression proteomics. Leukemia 16: 478-485.
- 112. Chin L, Hahn WC, Getz G, Meyerson M (2011) Making sense of cancer genomic data. Genes Dev 25: 534-555.
- Grondin RT, Scott RM, Smith ER (2009) Pediatric brain tumors. Adv Pediatr 56: 249-269.
- 114. Ward E, DeSantis C, Robbins A, Kohler B, Jemal A, (2014) Childhood and adolescent cancer statistics. CA Cancer J Clin 64: 83-103.
- 115. Bhatla T, Jones CL, Meyer JA, Vitanza NA, Raetz EA, et al. (2014) The biology of relapsed acute lymphoblastic leukemia: opportunities for therapeutic interventions. J Pediatr Hematol Oncol 36: 413-418.
- 116. Giordano TJ (2014) The cancer genome atlas research network: a sight to behold. Endocr Pathol 25: 362-365.
- 117. Greenman C, Stephens P, Smith R, Dalgliesh GL, Hunter C, et al. (2007) Patterns of somatic mutation in human cancer genomes. Nature 446: 153-158.
- 118. Lindqvist CM, Nordlund J, Ekman D, Johansson A, Moghadam BT, et al. (2015) The mutational landscape in pediatric acute lymphoblastic leukemia. deciphered by whole genome sequencing. Hum Mutat 36: 118-128.
- 119. Wang X, Liu Q, Zhang B (2014) Leveraging the complementary nature of RNA-Seq and shotgun proteomics data. Proteomics 14: 2676-2687.