

Research Article

Ratios of Calcium/Trace Elements as Prostate Cancer Markers

Vladimir Zaichick^{1*}, Sofia Zaichick²

¹Radionuclide Diagnostics Department, Medical Radiological Research Centre, Russia

²Laboratory of Dr. Gabriela Caraveo Piso, Feinberg School of Medicine, Northwestern University, Chicago, USA.

Corresponding author: Vladimir Zaichick, Radionuclide Diagnostics Department, Medical Radiological Research Centre Koroleva St- 4, Obninsk 249036, Kaluga Region, Russia, Tel: +7 (48439) 60289; Fax +7 (495) 956 1440; Email: vezai@obninsk.com

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Abstract

The aim of the study was to evaluate whether significant changes in the pro-static tissue levels of ratios Ca/trace element contents exist in the malignantly transformed prostate. Contents of Ca and 43 trace elements in normal (N, n=37), benign hypertrophic (BPH, n=32) and cancerous human prostate (PCa, n=60) were investigated, and ratios Ca/trace element contents were calculated. Measurements were performed using a combination of non-destructive and destructive methods: instrumental neutron activation analysis, inductively coupled plasma atomic emission spectrometry and inductively coupled plasma mass spectrometry. It was observed that the ratio to Ca of Ag, Al, Au, B, Be, Bi, Br, Ca, Cd, Ce, Co, Cr, Cs, Dy, Er, Fe, Gd, Hg, Ho, La, Li, Mn, Mo, Nb, Nd, Ni, Pb, Pr, Rb, Sb, Sc, Se, Sm, Sn, Tb, Th, Ti, Tl, Tm, U, Y, Yb, Zn, and Zr mass fraction were significantly lower in cancerous tissues than in normal and BPH prostate. Finally, we propose to use the Ca/Ag, Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios in a needle-biopsy core as an accurate tool to diagnose prostate cancer. Further studies on larger number of samples are required to confirm our findings and to investigate the impact of the trace element relationships on prostate cancer etiology.

Keywords: Benign Prostatic Hypertrophy; Inductively Coupled Plasma Atomic Emission Spectrometry; Inductively Coupled Plasma Mass Spectrometry; Neutron Activation Analysis; Prostate; Prostatic Carcinoma; Trace Elements; Trace Element Content Ratios.

Introduction

The prostate gland may be a source of many health problems in men past middle age, the most common Benign Prostatic Hyperplasia (BPH), and Prostatic Carcinoma (PCa). BPH is a non-cancerous enlargement of the prostate gland leading to obstruction of the urethra and can significantly impair quality of life. The prevalence of histological BPH is found in approximately 50-60% of males age 40-50, in over 70% at 60 years old and in greater than 90% of men over 70 [1,2]. In many Western industrialized countries, including North America, PCa is the most frequently diagnosed form of non-cutaneous malignancy in males. Except for lung cancer, PCa is the leading cause of death from cancer [3-8].

Although the etiology of BPH and PCa is unknown, some trace elements have been highlighted in the literature in relation to the development of these prostate diseases [9-29].

Trace elements have essential physiological functions such as maintenance and regulation of cell function and signaling, gene regulation, activation or inhibition of enzymatic reactions, neurotransmission, and regulation of membrane function. Essential or toxic (mutagenic, carcinogenic) properties of trace elements depend on tissue-specific need or tolerance [30]. Excessive accumulation, deficiency or an imbalance of the trace elements may disturb the cell functions and may result in cellular degeneration, death and malignant transformation [30].

In earlier reported studies [31-66] significant changes of trace element contents in hyperplastic and cancerous prostate in comparison with those in the normal prostatic tissue were observed. Moreover, a significant informative value of Ca content as a tumor marker for PCa diagnostics was shown by us [67,68].

Hence trace elements besides Ca can be used as tumor markers for distinguish between benign and malignant prostate.

Currently numbers of methods were applied for the measurement of chemical elements contents in samples of human tissue. Among these methods, the instrumental neutron activation analysis with high resolution spectrometry of short-lived radionuclides (INAA-SLR) and long-lived radionuclides (INAA-LLR) is a non-destructive and one of the most sensitive techniques. It allows measuring the trace element contents in few milligrams tissue without any treatment of sample. Analytical studies of the Ag, Br, Ca, Co, Cr, Fe, Hg, K, Mg, Mn, Na, Sb, Sc, Se, and Zn contents in normal, BPH and PCa tissue were done by us using INAA-SLR and INAA-LLR [14,15,20,27,28,53,54,60-62,64]. Nondestructive method of analysis avoids the possibility of changing the content of trace elements in the studied samples [69-72], which allowed for the first time to obtain reliable results. In particular, it was shown that the average mass fraction of Co, Cr, Hg, Sb, and Se in BPH were higher than normal levels [66]. In PCa tissues the mean values of Ag, Br, Cr, Fe, Hg, Mn, and Sb were higher while those of Ca, Co, Rb, Sc, and Zn were lower than in healthy prostatic tissue [60-65]. For example, the mean levels ($M \pm SEM$) of Ca mass fractions (mg/kg dry tissue) in normal, BPH, and cancerous prostate were 2428 ± 233 , 2032 ± 165 , and 676 ± 63 , respectively [61,64]. Obtained results formed the basis for a new method for differential diagnosis of BPH and PCa, the essence of which was to determine the ratios of chemical element contents changed in opposite directions during malignant transformation of prostate.

It is obvious that the most effective will be non-destructive analytical methods because they involve a minimal treatment of sample since the chances of significant loss or contamination would be decreased. However, the INAA allow only determining the mean mass fractions of 15-16 chemical elements in the samples of normal and cancerous prostate glands [14,15,20,27,28,60,66]. The combination of inductively coupled plasma atomic emission spectrometry (ICP-AES) and Inductively Coupled Plasma Mass Spectrometry (ICP-MS) are more power analytical tools than INAA [17,18] but sample digestion is a critical step in elemental analysis by these methods. In the present study all these analytical methods were used and the results obtained for some chemical elements by ICP-AES and ICP-MS were under the control of INAA data.

The present study had three aims. The main objective was to obtain reliable results about the 44 chemical elements: Ag, Al, Au, B, Be, Bi, Br, Ca, Cd, Ce, Co, Cr, Cs, Dy, Er, Fe, Gd, Hg, Ho, La, Li, Mn, Mo, Nb, Nd, Ni, Pb, Pr, Rb, Sb, Sc, Se, Sm, Sn, Tb, Th, Ti, Tl, Tm, U, Y, Yb, Zn, and Zr contents in intact prostate of healthy men as well as in the prostate gland of patients with BPH and PCa using INAA and ICP methods. The second aim was to

calculate Ca/trace element content ratios and compare the levels of these ratios in normal, hyperplastic, and cancerous prostate. The third and final aim was to evaluate the ratios of Ca/trace element contents for diagnosis of prostate cancer. All studies were approved by the Ethical Committees of the Medical Radiological Research Centre, Obninsk.

Material and Methods

Samples

The patients studied (n=92) were hospitalized in the Urological Department of the Medical Radiological Research Centre (Obninsk, Russia). All of them were European-Caucasian, citizens of Moscow and Obninsk (a small city in a non-industrial region 105 km south-west of Moscow). Transrectal puncture biopsy of suspicious indurated regions of the prostate was performed for every patient, to permit morphological study of prostatic tissue at these sites and to estimate their chemical element contents. The diagnosis of all patients has been confirmed by clinical and morphological results obtained during studies of biopsy and respected materials. The age of 32 patients with BPH ranged from 56 to 78 years, the mean being 66 ± 6 ($M \pm SD$) years. The 60 patients aged 40-79 suffered from PCa (stage T1-T4). Their mean age was 65 ± 10 ($M \pm SD$) years.

Intact prostates (N) were removed at necropsy from 37 men aged 41-87 who had died suddenly. All deceased were European-Caucasian, citizens of Moscow. Their mean age was 55 ± 11 ($M \pm SD$) years. Majority of deaths were due to trauma. Tissue samples were collected from the peripheral zone of dorsal and lateral lobes of their prostates, within 2 days of death and then the samples were divided into two portions. One was used for morphological study while the other was intended for chemical element analysis. A histological examination was used to control the age norm conformity, as well as to confirm the absence of microadenomatosis and latent cancer [14,15,20,28].

Sample Preparation

All tissue samples were divided into two portions. One was used for morphological study while the other was intended for trace element analysis. The samples intended for trace element analysis were weighed, freeze-dried and homogenized. The sample weighing about 10 mg (for biopsy materials) and 50-100 mg (for respected materials) was used for Ca measurement by INAA-SLR. The samples for INAA-SLR were sealed separately in thin polyethylene films washed with acetone and rectified alcohol before using. The sealed samples were placed in labeled polyethylene ampoules. After NAA-SLR investigation, the prostate samples were taken out from the polyethylene ampoules and used for trace element measurement by INAA-LLR. The samples for INAA-LLR were

wrapped separately in a high-purity aluminum foil washed with double rectified alcohol beforehand and placed in a nitric acid-washed quartz ampoule.

After NAA-LLR investigation, the prostate samples were taken out and used for ICP methods. The samples were decomposed in autoclaves; 1.5 mL of concentrated HNO_3 (nitric acid at 65 %, maximum (max) of 0.0000005 % Hg; GR, ISO, Merck) and 0.3 mL of H_2O_2 (pure for analysis) were added to prostate tissue samples, placed in one-chamber autoclaves (Ancon-AT2, Ltd., Russia) and then heated for 3 h at 160–200°C. After autoclaving, they were cooled to room temperature and solutions from the decomposed samples were diluted with deionized water (up to 20 mL) and transferred to the plastic measuring bottles. Simultaneously, the same procedure was performed in autoclaves without tissue samples (only $\text{HNO}_3+\text{H}_2\text{O}_2+\text{deionized water}$), and the resultant solutions were used as control samples.

Instrumentation and Methods

INAA

A horizontal channel equipped with the pneumatic rabbit system of the WWR-C research nuclear reactor was applied to determine the mass fraction of Ca by INAA-SLR. The neutron flux in the channel was $1.7 \times 10^{13} \text{n cm}^{-2}\text{s}^{-1}$. Ampoules with prostate samples, biological synthetic standards [73], intralaboratory-made standards, and Certified Reference Material (CRM) were put into polyethylene rabbits and irradiated separately for 180 s. Copper foils were used to assess neutron flux by the detection of the 511 keV gamma line of [64] Cu from reaction [63] $\text{Cu}(\text{n},\gamma)$ [64] Cu. One minute after the irradiation, the measurement of gamma ray activities of the reaction products for each sample was done using gamma ray spectrometer. The duration of each measurement was 10 min. The detector to sample distance was from 5 to 15 cm subject to pulse counting rate.

A vertical channel of a nuclear reactor was applied to determine the trace element mass fractions by NAA-LLR. The quartz ampoule with prostate samples and certified reference materials was soldered, positioned in a transport aluminum container and exposed to a 24-hour neutron irradiation in a vertical channel with a neutron flux of $1.3 \times 10^{13} \text{n cm}^{-2}\text{s}^{-1}$. Ten days after the irradiation, samples were reweighed and repacked. Over a period from 10 to 30 days after irradiation, the samples were measured for the gamma ray activities of the reaction products by using gamma ray spectrometer. The duration of measurements was from 20 min to 10 hours subject to pulse counting rate. The detector to sample distance was 0 cm.

The gamma ray spectrometer used for NAA-SLR and NAA-LLR included the $100 \text{ cm}^3 \text{Ge(Li)}$ detector and on-line computer-

based multichannel analyzer. The spectrometer provided a resolution of 1.9 keV at the 1332 keV gamma line of [60] Co. Information detailing with the NAA-SLR and NAA-LLR methods used and other details of the analysis was presented in our previous publications [14,15].

ICP-AES and ICP-MS

Aliquots of aqueous solutions were used to determine the Ca mass fractions by ICP-AES using the Spectrometer ICAP-61 (Thermo Jarrell Ash, USA). Integration time of the spectrum during measurement was 5 s. The determination of the Ca content in aqueous solutions was made by the quantitative method using calibration solutions (High Purity Standards, USA) of 0.5 and 10 mg/L. The calculations of the Ca content in the probe were carried out using software of a spectrometer (ThermoSPEC, version 4.1). An ICP-MS Thermo-Fisher "X-7" Spectrometer (Thermo Electron, USA) was used to determine the content of trace elements by ICP-MS. The element concentrations in aqueous solutions were determined by the quantitative method using multi elemental calibration solutions ICP-MS-68A and ICP-AM-6-A produced by High-Purity Standards (Charleston, SC29423, USA). Indium was used as an internal standard in all measurements.

Information detailing with the ICP-AES and ICP-MS methods used and other details of the analysis was presented in our previous publications [17,18].

Certified Reference Materials

For quality control, ten subsamples of the certified reference material IAEA H-4 (Animal muscle) from the International Atomic Energy Agency were used. In addition, five sub-samples INCT-SBF-4 Soya Bean Flour, INCT-TL-1 Tea Leaves and INCT-MPH-2 Mixed Polish Herbs from the Institute of Nuclear Chemistry and Technology (INCT, Warszawa, Poland) were analyzed simultaneously with the investigated prostate tissue samples. All samples of CRM were treated in the same way as the prostate samples. Detailed results of this quality assurance program were presented in earlier publications [14,15,17,18].

Computer Programs and Statistic

A dedicated computer program for INAA mode optimization was used [74]. All prostate samples for NAA-SLR and INAA-LLR were prepared in duplicate and mean values of chemical element contents were used in final calculation. For elements investigated by NAA-SLR, INAA-LLR, ICP-AES, and ICP-MS the mean of all results was used. Using the Microsoft Office Excel software Ca/trace element contents for each trace element in every sample were calculated. Then arithmetic mean, standard deviation, and standard error of mean were calculated for ratios of Ca/trace element mass

fraction in normal, benign hyperplastic and cancerous prostate tissue. The difference in the results between BPH and Norm, PCa and Norm as well as PCA and BPH was evaluated by parametric Student's t-test and non-parametric Wilcoxon-Mann-Whitney U-test. Values of $p<0.05$ is considered to be statistically significant. For the construction of "individual data sets for Ca/trace element mass fraction ratios in normal, benign hypertrophic and cancerous prostate" diagrams the Microsoft Office Excel software was also used.

Results

Table 1 depicts mean values \pm standard error of mean ($M\pm SEM$) of the ratio to Ca of Ag, Al, Au, B, Be, Bi, Br, Ca, Cd, Ce, Co, Cr, Cs, Dy, Er, Fe, Gd, Hg, Ho, La, Li, Mn, Mo, Nb, Nd, Ni, Pb, Pr, Rb, Sb, Sc, Se, Sm, Sn, Tb, Th, Ti, Tl, Tm, U, Y, Yb, Zn, and Zr mass fraction in normal, benign hypertrophic and cancerous prostate.

Ratio of chemical elements	Symbols	Prostatic tissue		
		N	BPH	PCa
		41-87 year (n=37)	56-78 year (n=32)	40-79 year (n=60)
Calcium/Silver	Ca/Ag	107037 \pm 15764	117550 \pm 34515	4164 \pm 1611
Calcium/Aluminum	Ca/Al	103 \pm 21	101 \pm 18	5.24 \pm 1.96
Calcium/Gold	Ca/Au	1170772 \pm 216470	1126774 \pm 158450	66619 \pm 34235
Calcium/Boron	Ca/B	4320 \pm 805	1550 \pm 191	119 \pm 587
Calcium/Berillium	Ca/Be	3096156 \pm 413429	2224935 \pm 159071	94738 \pm 39812
Calcium/Bismuth	Ca/Bi	532698 \pm 114578	93934 \pm 41193	7501 \pm 6138
Calcium/Bromine	Ca/Br	104 \pm 13	75.8 \pm 13.1	11.2 \pm 4.2
Calcium/Cadmium	Ca/Cd	3085 \pm 455	3753 \pm 732	2002 \pm 212
Calcium/Cerium	Ca/Ce	144087 \pm 28909	191735 \pm 31186	8862 \pm 2222
Calcium/Cobalt	Ca/Co	69329 \pm 11034	42314 \pm 4982	13669 \pm 1071
Calcium/Cromium	Ca/Cr	14516 \pm 6572	2169 \pm 218	191 \pm 41
Calcium/Cesium	Ca/Cs	94882 \pm 17335	92843 \pm 9485	22723 \pm 6474
Calcium/Dysprosium	Ca/Dy	1733952 \pm 444593	1685620 \pm 327920	161389 \pm 47689
Calcium/Erbium	Ca/Er	2782727 \pm 557202	3989832 \pm 845199	296667 \pm 64924
Calcium/Iron	Ca/Fe	25.8 \pm 3.8	15.8 \pm 1.7	4.88 \pm 0.57
Calcium/Gadolinium	Ca/Gd	1489869 \pm 334486	1726552 \pm 327682	118454 \pm 35677
Calcium/Mercury	Ca/Hg	78186 \pm 11882	9944 \pm 1259	4445 \pm 2330
Calcium/Holmium	Ca/Ho	7482530 \pm 1547065	8358623 \pm 1644445	453653 \pm 88284
Calcium/Lanthanum	Ca/La	105705 \pm 19452	128328 \pm 16885	7340 \pm 3488
Calcium/Lithium	Ca/Li	79700 \pm 10834	71782 \pm 11904	5847 \pm 2025
Calcium/Manganese	Ca/Mn	1980 \pm 297	1789 \pm 186	181 \pm 66
Calcium/Molybdenum	Ca/Mo	11809 \pm 1417	12394 \pm 1087	2585 \pm 705
Calcium/Niobium	Ca/Nb	8481196 \pm 152944	952286 \pm 168195	168030 \pm 22370
Calcium/Neodymium	Ca/Nd	320718 \pm 67884	403461 \pm 65435	21961 \pm 4822
Calcium/Nickel	Ca/Ni	1916 \pm 626	1028 \pm 179	123 \pm 28
Calcium/Lead	Ca/Pb	3774 \pm 724	4461 \pm 756	556 \pm 149
Calcium/Praseodymium	Ca/Pr	1263853 \pm 269288	2231480 \pm 735010	112525 \pm 35218
Calcium/Rubidium	Ca/Rb	219 \pm 36	139 \pm 12	81.6 \pm 7.4
Calcium/Antimony	Ca/Sb	121969 \pm 24740	49028 \pm 20319	2784 \pm 556
Calcium/Scandium	Ca/Sc	174958 \pm 51707	76931 \pm 10995	49945 \pm 6858
Calcium/Selenium	Ca/Se	3604 \pm 500	2362 \pm 296	895 \pm 168

Calcium/Samarium	Ca/Sm	1695658±438262	2939100±810027	142073±37596
Calcium/Tin	Ca/Sn	15885±2313	30043±4945	1242±490
Calcium/Terbium	Ca/Tb	12456254±2477137	14659058±2058440	897422±223585
Calcium/Thorium	Ca/Th	1703628±375869	1499284±275749	43707±21300
Calcium/Titanium*	Ca/Ti*	2079±456	1587±223	144±67
Calcium/Thallium	Ca/Tl	2870569±627543	1464103±251751	124174±74903
Calcium/Thulium	Ca/Tm	17965352±4197256	17684696±3565514	1543386±608859
Calcium/Uranium	Ca/U	1122953±182815	2061625±434930	130793±22073
Calcium/Yttrium	Ca/Y	414434±116105	385834±74931	23034±3863
Calcium/Ytterbium	Ca/Yb	3755556±862110	4778651±1310265	435607±60354
Calcium/Zinc	Ca/Zn	4.00±0.93	1.72±0.21	5.38±0.47
Calcium/Zirconium	Ca/Zr	135701±31300	61766±18949	853±238

M - arithmetic mean, SEM - standard error of mean, NS - not significant difference.

*Titanium tools were used for sampling and sample preparation.

Table 1: Comparison of mean values (M±SEM) of the calcium/trace element mass fraction ratios in normal(N), Benign Hypertrophic (BPH) and Cancerous Prostate (PCa).

The ratios of means and the difference between mean values of the ratio to Ca of Ag, Al, Au, B, Be, Bi, Br, Ca, Cd, Ce, Co, Cr, Cs, Dy, Er, Fe, Gd, Hg, Ho, La, Li, Mn, Mo, Nb, Nd, Ni, Pb, Pr, Rb, Sb, Sc, Se, Sm, Sn, Tb, Th, Ti, Tl, Tm, U, Y, Yb, Zn, and Zr mass fraction in normal, benign hypertrophic and cancerous prostate are presented in Table 2.

	BPH and Normal (N)			PCa and Normal (N)			PCa and BPH		
	Ratio	p≤	p	Ratio	p≤	p	Ratio	p≤	p
	BPH/N	t-test	U-test	PCa/N	t-test	U-test	PCa/BPH	t-test	U-test
Ca/Ag	1.1	0.785	>0.05	0.039	0.000001	≤0.01	0.035	0.0082	≤0.01
Ca/Al	0.98	0.936	>0.05	0.051	0.000093	≤0.01	0.052	0.00047	≤0.01
Ca/Au	0.96	0.871	>0.05	0.057	0.000074	≤0.01	0.059	0.000043	≤0.01
Ca/B	0.36	0.0028	≤0.01	0.028	0.000036	≤0.01	0.077	0.00004	≤0.01
Ca/Be	0.72	0.059	≤0.05	0.031	0.000001	≤0.01	0.043	0.000001	≤0.01
Ca/Bi	0.18	0.0012	≤0.01	0.014	0.00013	≤0.01	0.08	0.064	≤0.05
Ca/Br	0.73	0.14	>0.05	0.108	0.000001	≤0.01	0.148	0.00052	≤0.01
Ca/Cd	1.22	0.448	>0.05	0.649	0.039	≤0.01	0.533	0.041	≤0.01
Ca/Ce	1.33	0.272	>0.05	0.062	0.000012	≤0.01	0.046	0.000016	≤0.01
Ca/Co	0.61	0.033	≤0.05	0.197	0.000038	≤0.01	0.323	0.000016	≤0.01
Ca/Cr	0.15	0.074	≤0.01	0.013	0.041	≤0.01	0.088	0.000006	≤0.01
Ca/Cs	0.98	0.918	>0.05	0.239	0.000051	<0.01	0.245	0.00001	≤0.01
Ca/Dy	0.97	0.931	>0.05	0.093	0.0019	≤0.01	0.096	0.000088	≤0.01
Ca/Er	1.43	0.248	>0.05	0.107	0.0002	≤0.01	0.074	0.0014	≤0.01
Ca/Fe	0.61	0.021	≤0.05	0.189	0.000009	≤0.01	0.309	0.000063	≤0.01
Ca/Gd	1.16	0.617	>0.05	0.08	0.00048	≤0.01	0.069	0.0006	≤0.01
Ca/Hg	0.13	6.00E-06	≤0.05	0.057	0.000002	≤0.01	0.447	0.08	≤0.05
Ca/Ho	1.12	0.701	>0.05	0.061	0.00015	≤0.01	0.054	0.00071	≤0.01
Ca/La	1.21	0.387	>0.05	0.069	0.000038	≤0.01	0.057	0.000041	≤0.01

Ca/Li	0.9	0.627	>0.05	0.073	0.000001	≤0.01	0.081	0.00023	≤0.01
Ca/Mn	0.9	0.59	>0.05	0.091	0.000004	≤0.01	0.101	0.000002	≤0.01
Ca/Mo	1.05	0.746	>0.05	0.219	0.000004	≤0.01	0.209	0.000003	≤0.01
Ca/Nb	0.11	0.651	>0.05	0.02	0.00021	≤0.01	0.176	0.00087	≤0.01
Ca/Nd	1.26	0.388	>0.05	0.068	0.00025	≤0.01	0.054	0.00016	≤0.01
Ca/Ni	0.54	0.184	>0.05	0.064	0.009	≤0.01	0.12	0.00047	≤0.01
Ca/Pb	1.18	0.517	>0.05	0.147	0.00019	≤0.01	0.125	0.00038	≤0.01
Ca/Pr	1.77	0.239	>0.05	0.089	0.00032	≤0.01	0.05	0.016	≤0.01
Ca/Rb	0.63	0.041	≤0.01	0.373	0.00069	≤0.01	0.587	0.00071	≤0.01
Ca/Sb	0.4	0.029	≤0.01	0.023	0.000055	≤0.01	0.057	0.046	≤0.01
Ca/Sc	0.44	0.085	>0.05	0.285	0.032	≤0.01	0.649	0.083	≤0.05
Ca/Se	0.66	0.04	≤0.05	0.248	0.000018	≤0.01	0.379	0.00073	≤0.01
Ca/Sm	1.73	0.196	>0.05	0.084	0.0018	≤0.01	0.048	0.0062	≤0.01
Ca/Sn	1.89	0.021	≤0.01	0.078	0.000001	≤0.01	0.041	0.00016	≤0.01
Ca/Tb	1.18	0.499	>0.05	0.072	0.00012	≤0.01	0.061	0.000051	≤0.01
Ca/Th	0.88	0.664	>0.05	0.026	0.00022	≤0.01	0.029	0.00035	≤0.01
Ca/Ti*	0.76	0.342	>0.05	0.069	0.00038	≤0.01	0.091	0.000054	≤0.01
Ca/Tl	0.51	0.047	≤0.05	0.043	0.00025	≤0.01	0.085	0.00039	≤0.01
Ca/Tm	0.98	0.96	>0.05	0.086	0.00078	≤0.01	0.087	0.0011	≤0.01
Ca/U	1.84	0.067	≤0.05	0.116	0.000016	≤0.01	0.063	0.0013	≤0.01
Ca/Y	0.93	0.837	>0.05	0.056	0.0025	≤0.01	0.06	0.00068	≤0.01
Ca/Yb	1.27	0.522	>0.05	0.116	0.00088	≤0.01	0.091	0.0078	≤0.01
Ca/Zn	0.43	0.024	≤0.05	1.345	0.195	>0.05	3.128	0.000006	≤0.01
Ca/Zr	0.46	0.052	≤0.05	0.006	0.00028	≤0.01	0.014	0.011	≤0.01

t-test - Student's t-test, U-test - Wilcoxon-Mann-Whitney U-test, Bold significant differences

Table 2: Ratio of means and the difference between mean values of the Ca mass fraction/ trace element mass fraction ratios in normal(N), Benign Hypertrophic (BPH) and Cancerous Prostate (PCa).

Individual data sets for Ca/Ag, Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios in all

investigated samples of normal, benign hypertrophic and cancerous prostate, respectively, are shown in Figure 1.

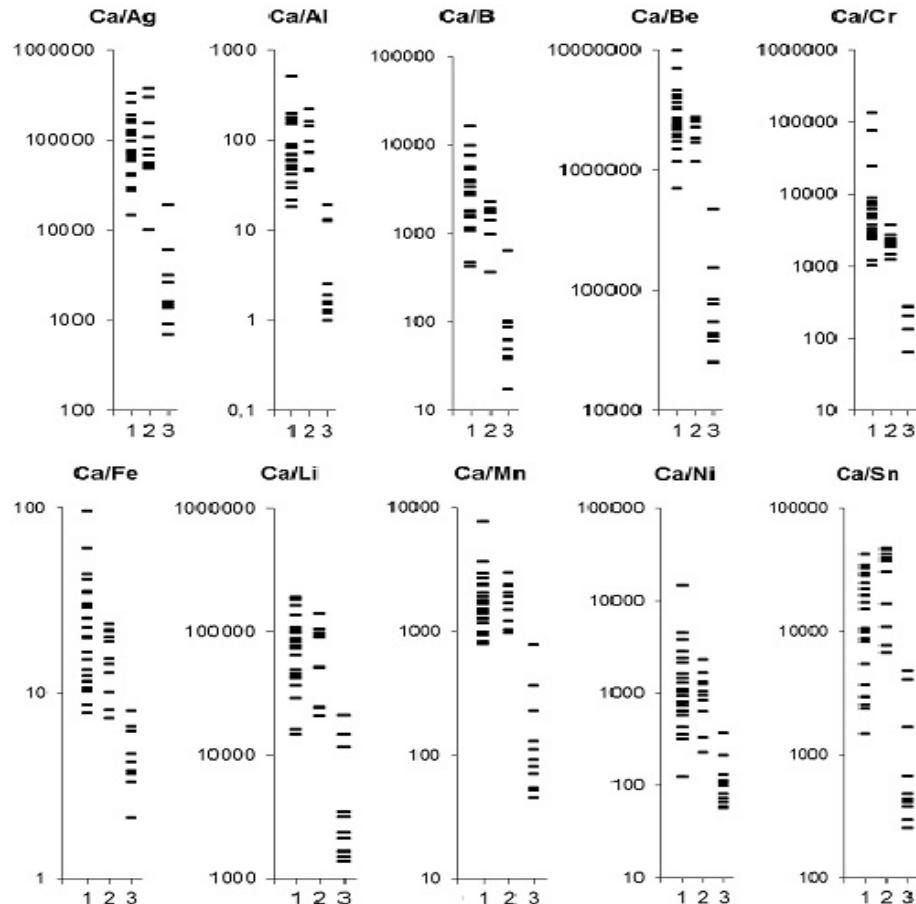


Figure 1: Individual data sets for Ca/Ag, Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios in samples of normal (1), benign hypertrophic (2) and cancerous (3) prostate.

Table 3 contains parameters of the importance (sensitivity, specificity and accuracy of Ca/Ag, Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios for the diagnosis of PCa calculated in this work.

Mass fraction ratio or their multiplication	Upper limit for PCa	Sensitivity %	Specificity %	Accuracy %
Ca/Ag	8000	91±9	100-3	98±2
Ca/Al	17	91±9	100-3	98±2
Ca/B	400	90±10	100-3	98±2
Ca/Be	500000	100-9	100-3	100-2
Ca/Cr	300	100-20	100-3	100-3
Ca/Fe	7	90±10	100-3	98±2
Ca/Li	14600	91±9	100-3	98±2

Ca/Mn	780	100-9	100-3	100-2
Ca/Ni	215	91±9	97±3	96±3
Ca/Sn	2000	82±12	97±3	94±4

Table 3: Parameters of the importance (sensitivity, specificity and accuracy) of Ca/Ag, Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios for the diagnosis of PCa (an estimation is made for “PCa or normal and BPH prostate”).

Discussion

As was shown by us [14,15,17,18], the use of CRM IAEA H-4, INCT-SBF-4 Soya Bean Flour, INCT-TL-1 Tea Leaves, and INCT-MPH-2 Mixed Polish Herbs as certified reference materials for the analysis of samples of prostate tissue can be seen as quite acceptable. Good agreement of the Ag, Al, Au, B, Be, Bi, Br, Ca, Cd, Ce, Co, Cr, Cs, Dy, Er, Fe, Gd, Hg, Ho, La, Li, Mn, Mo, Nb, Nd, Ni, Pb, Pr, Rb, Sb, Sc, Se, Sm, Sn, Tb, Th, Ti, Tl, Tm, U, Y, Yb,

Zn and Zr contents analyzed by NAA-SLR, NAA-LLR, ICP-AES, and ICP-MS with the certified data of reference materials indicates an acceptable accuracy of the results obtained in the study of trace elements of the prostate samples presented in Tables 1 and 2.

The mean values and all selected statistical parameters were calculated for 43 ratios of Ca/trace element contents (Table 1). The mass fraction of Ca and 43 trace elements were measured in all, or a major portion of normal prostate samples. The masses of BPH and PCa samples varied very strong from a few milligrams (sample from needle biopsymaterial) to 100 mg (sample from respected material). Therefore, in BPH and PCa prostates mass fraction ratios of Ca/trace element content were determined in 22 samples (11 BPH and 11 PCa samples, respectively).

From Table 2, it is observed that in benign hypertrophic tissues the Ca/Ag, Ca/Al, Ca/Au, Ca/Br, Ca/Cd, Ca/Ce, Ca/Cs, Ca/Dy, Ca/Er, Ca/Gd, Ca/Ho, Ca/La, Ca/Mo, Ca/Nb, Ca/Nd, Ca/Ni, Ca/Pb, Ca/Pr, Ca/Sc, Ca/Sm, Ca/Tb, Ca/Th, Ca/Ti, Ca/Tm, Ca/Y, Ca/Yb, and Ca/Zr mass fraction ratios not differ from normal levels, but the mass fraction ratios of Ca/Sn and Ca/U are higher, while the mass fraction ratios of Ca/B, Ca/Be, Ca/Bi, Ca/Co, Ca/Cr, Ca/Fe, Ca/Hg, Ca/Rb, Ca/Sb, Ca/Se, Ca/Tl, Ca/Zn, and Ca/Zr are significantly lower. In cancerous tissue the all Ca/trace element mass fraction ratios investigated in the study are significantly lower than in BPH and normal prostate with the exception of Ca/Zn ratio.

Analysis of the mass fraction ratios for trace element in prostate tissue could become a powerful diagnostic tool. To a large extent, the resumption of the search for new methods for early diagnosis of PCa was due to experience gained in a critical assessment of the limited capacity of the prostate specific antigen (PSA) serum test [75,76]. In addition to the PSA serum test and morphological study of needle-biopsy cores of the prostate, the development of other highly precise testing methods seems to be very useful. Experimental conditions of the present study were approximated to the hospital conditions as closely as possible. In BPH and PCa cases we analyzed a part of the material obtained from a puncture transrectal biopsy of the indurate site in the prostate. Therefore, our data allow us to evaluate adequately the importance of Ca/trace element mass fraction ratios for the diagnosis of PCa. As is evident from Table 2 and, particularly, from individual data sets (Figure 1), the Ca/Ag, Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios are potentially the most informative test for a differential diagnosis. For example, if 8000 is the value of Ca/Ag mass fraction ratio assumed to be the upper limit for PCa (Figure 1) and an estimation is made for "PCa or intact and BPH tissue", the following values are obtained:

$$\text{Sensitivity} = \{\text{True Positives (TP)} / [\text{TP} + \text{False Negatives (FN)}]\} \cdot 100\% = 91 \pm 9\%.$$

$$\text{Specificity} = \{\text{True Negatives (TN)} / [\text{TN} + \text{False Positives (FP)}]\} \cdot 100\% = 100 - 3\%;$$

$$\text{Accuracy} = [(\text{TP} + \text{TN}) / (\text{TP} + \text{FP} + \text{TN} + \text{FN})] \cdot 100\% = 98 \pm 2\%.$$

The number of people (samples) examined was taken into account for calculation of confidence intervals [77]. In other words, if Ca/Ag mass fraction ratio in a prostate biopsy sample is lower than 8000, one could diagnose a malignant tumor with an accuracy 98±2%. Thus, using the Ca/Ag mass fraction ratio-test makes it possible to diagnose cancer in 91±9% cases (sensitivity). The same way parameters of the importance (sensitivity, specificity and accuracy) of Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios for the diagnosis of PCa were calculated (Table 3).

Conclusion

The combination of nondestructive INAA and destructive ICP methods is satisfactory analytical tool for the precise determination of Ca and 43 trace element mass fractions in the tissue samples of normal, BPH and carcinomatous prostate glands. The sequential application of these methods allowed precise quantitative determinations of mean mass fraction of Ag, Al, Au, B, Be, Bi, Br, Ca, Cd, Ce, Co, Cr, Cs, Dy, Er, Fe, Gd, Hg, Ho, La, Li, Mn, Mo, Nb, Nd, Ni, Pb, Pr, Rb, Sb, Sc, Se, Sm, Sn, Tb, Th, Ti, Tl, Tm, U, Y, Yb, Zn and Zr. It was observed that the ratio to Ca of Ag, Al, Au, B, Be, Bi, Br, Ca, Cd, Ce, Co, Cr, Cs, Dy, Er, Fe, Gd, Hg, Ho, La, Li, Mn, Mo, Nb, Nd, Ni, Pb, Pr, Rb, Sb, Sc, Se, Sm, Sn, Tb, Th, Ti, Tl, U, Y, Yb, and Zr mass fraction were significantly lower in cancerous tissues than in normal and BPH prostate. Finally, we propose to use the Ca/Ag, Ca/Al, Ca/B, Ca/Be, Ca/Cr, Ca/Fe, Ca/Li, Ca/Mn, Ca/Ni, and Ca/Sn mass fraction ratios in a needle-biopsy core as an accurate tool to diagnose prostate cancer. Further studies on larger number of samples are required to confirm our findings and to investigate the impact of the trace element relationships on prostate cancer etiology.

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References

1. Roehrborn C, McConnell J (2002) Etiology, pathophysiology, epidemiology and natural history of benign prostatic hyperplasia. In: Camp-

bell's Urology (8th edn). Saunders, Philadelphia: 1297-1336.

- 2. Lepor H (2005) Pathophysiology of benign prostatic hyperplasia in the aging male population. *Rev Urol* 7(Suppl 4): S3-S12.
- 3. Oliver SE, Gunnell D, Donovan JL (2000) Comparison of trends in prostate-cancer mortality in England and Wales and the USA. *Lancet* 355:1788-1789.
- 4. Kumar RJ, Barqawi AB, Crawford ED (2004) Epidemiology of prostate cancer. *Business Briefing: US Oncology Review*: 1-6.
- 5. Maddams J, Brewster D, Gavin A, Steward J, Elliott J, et al. (2009) Cancer prevalence in the United Kingdom: estimates for 2008. *Br J Cancer* 101: 541-547.
- 6. Möller T, Anderson H, Aarelid T, Hakulinen T, Storm H, et al. (2003) Cancer prevalence in Northern Europe: the EUROPREVAL study. *Ann Oncol* 14: 946-957.
- 7. De Angelis R, Grande E, Inghelmann R, Francisci S, Micheli A, et al. (2007) Cancer prevalence estimates in Italy from 1970 to 2010. *Tumori* 93: 392-397.
- 8. Waalkes MP, Rehm S (1994) Cadmium and prostate cancer. *J Toxicol Environ Health* 43: 251-269.
- 9. Zaichick V, Zaichick S (1999) Role of zinc in prostate cancerogenesis. In: Anke M, et al, editors. *Mengen und Spurenelemente*. 19. Arbeitstagung. Friedrich-Schiller-Universität. Jena: 104-115.
- 10. Platz EA, Helzlsouer KJ (2001) Selenium, zinc, and prostate cancer. *Epidemiol Rev* 23: 93-101.
- 11. Zaichick V (2004) INAA and EDXRF applications in the age dynamics assessment of Zn content and distribution in the normal human prostate. *J Radioanal Nucl Chem* 262: 229-234.
- 12. Gray MA, Centeno JA, Slaney DP, Ejnik JW, Todorov T, et al. (2005) Environmental exposure to trace elements and prostate cancer in three New Zealand ethnic groups. *Int J Environ Res Public Health* 2: 374-384.
- 13. Zaichick S, Zaichick V (2011) INAA application in the age dynamics assessment of Br, Ca, Cl, K, Mg, Mn, and Na content in the normal human prostate. *J Radioanal Nucl Chem* 288: 197-202.
- 14. Zaichick S, Zaichick V (2011) The effect of age on Ag, Co, Cr, Fe, Hg, Sb, Sc, Se, and Zn contents in intact human prostate investigated by neutron activation analysis. *Appl Radiat Isot* 69: 827-833.
- 15. Zaichick S, Zaichick V (2011) The Br, Fe, Rb, Sr, and Zn content and interrelation in intact and morphologic normal prostate tissue of adult men investigated by energy dispersive X-ray fluorescent analysis. *X-Ray Spectrom* 40: 464-469.
- 16. Zaichick V, Nosenko S, Moskvinal (2012) The effect of age on 12 chemical element contents in intact prostate of adult men investigated by inductively coupled plasma atomic emission spectrometry. *Biol Trace Elem Res* 147: 49-58.
- 17. Zaichick S, Zaichick V, Nosenko S, Moskvina I (2012) Mass Fractions of 52 Trace Elements and Zinc Trace Element Content Ratios in Intact Human Prostates Investigated by Inductively Coupled Plasma Mass Spectrometry. *Biol Trace Elem Res* 149: 171-183.
- 18. Zaichick V, Zaichick S (2014) Age-related histological and zinc content changes in adult non hyper plastic prostate glands. *Age* 36: 167-181.
- 19. Zaichick V, Zaichick S (2014) INAA application in the assessment of chemical element mass fractions in adult and geriatric prostate glands. *Appl Radiat Isot* 90: 62-73.
- 20. Zaichick V, Zaichick S (2014) Determination of trace elements in adults and geriatric prostate combining neutron activation with inductively coupled plasma atomic emission spectrometry. *Open Journal of Biochemistry* 1: 16-33.
- 21. Zaichick V, Zaichick S (2014) Use of INAA and ICP-MS for the assessment of trace element mass fractions in adult and geriatric prostate. *J Radioanal Nucl Chem* 301: 383-397.
- 22. Zaichick V (2015) The variation with age of 67 macro- and microelement contents in non hyperplastic prostate glands of adult and elderly males investigated by nuclear analytical and related methods. *Biol Trace Elem Res* 168: 44-60.
- 23. Zaichick V, Zaichick S (2015) Dietaryintake of minerals and prostate cancer: insights into problem based on the chemical element contents in the prostate gland. *J Aging Res Clin Practice* 4: 164-171.
- 24. Zaichick V, Zaichick S (2015) Global contamination from uranium: insights into problem based on the uranium content in the human prostate gland. *J Environ Health Sci* 1: 1-5.
- 25. Zaichick V, Zaichick S (2016) Variations in concentration and distribution of several androgen-dependent and -independent trace elements in non-hyperplastic prostate gland tissue throughout adulthood. *J Androl Gynaecol* 4: 1-10.
- 26. Zaichick V, Zaichick S (2016) Age-related changes in concentration and histological distribution of Br, Ca, Cl, K, Mg, Mn, and Na in non hyperplastic prostate of adults. *European Journal of Biology and Medical Science Research* 4: 31-48.
- 27. Zaichick V, Zaichick S (2016)Variations in concentration and histological distribution of Ag, Co, Cr, Fe, Hg, Rb, Sb, Sc, Se, and Zn in nonhyperplastic prostate gland throughout adulthood. *Jacobs Journal of Cell and Molecular Biology* 2: 11.
- 28. Zaichick V, Zaichick S (2016) Age-relatedchangesinconcentrationandhistological distribution of 54 trace elements in nonhyperplastic prostate of adults. *Int Arch Urol Complic* 2: 19.
- 29. Zaichick V (2006) Medical elementology as a new scientific discipline. *J Radioanal Nucl Chem* 269: 303-309.
- 30. Stitich SR (1957) Trace elements in human tissue. I. A semi-quantitative spectrographicsurvey. *Biochem J* 67: 97-103.
- 31. Tipton IH, Cook MJ (1963) Trace elements in human tissue. Part II. Adult subjects from the United States. *Health Phys* 9: 103-145.
- 32. Györkey F, Min K-W, Huff JA, Györkey P (1967) Zinc and magnesium in human prostate gland: normal, hyperplastic, and neoplastic. *Cancer Res* 27: 1349-1353.
- 33. Sangen H (1967) The influence of the trace metals upon the aconitase activity in human prostate glands. *Jap J Uro* 158: 1146-1159.
- 34. Schneider H-J, Anke M, Holm W (1970) The inorganic components of testicle, epididymis, seminal vesicle, prostate and ejaculate of young men. *Int Urol Nephrol* 2: 419-427.
- 35. Hienzsc hE, Schneider H-J, Anke M (1970) Vergleichende Untersuchungen zum Mengen- und Spurenelementgehalt der normalen Prostata, des Prostataadenoms und des Prostatakarzinoms. *Zeitschrift für Urologie und Nephrologie* 63: 543-546.
- 36. Soman SD, Joseph KT, Raut SJ, Mulay GD, Parameswaran M, et al.

(1970) Studies of major and trace element content in human tissues. *Health Phys* 19: 641-656.

- 37. Forssen A (1972) Inorganic elements in the human body. I. occurrence of Ba, Br, Ca, Cd, Cs, Cu, K, Mn, Ni, Sn, Sr, Y and Zn in the human body. *Annales medicinae Experimental isetBiologie* 50: 99-162.
- 38. Dhar NK, Goel TC, Dube PC, Chowdhury AR, Kar AB (1973) Distribution and concentration of zinc in the subcellular fractions of benign hyperplastic and malignant neoplastic human prostate. *Exp Mol Pathol* 19: 139-142.
- 39. Jafa A, Mahendra NM, Chowdhury AR, Kamboj VP (1980) Trace elements in prostatic tissue and plasma in prostatic diseases of man. *Indian J Cancer* 17: 34-37.
- 40. Marezynska A, Kulpa J, Lenko J (1983) The Concentration of zinc in relation to fundamental elements in the diseases human prostate. *Int Urol Nephrol* 15: 257-265.
- 41. Hienzsch E, Schneider H-J, Anke M (1991) Vergleichende Untersuchungen zum Mengen- und Spurenelementgehalt der normalen Prostata, des Prostataadenoms und des Prostatakarzinoms. *Z Urol Nephrol* 163: 543-546.
- 42. Picurelli L, Olcina PV, Roig MD, Ferrer J (1991) Determination of Fe, Mg, Cu, and Zn in normal and pathological prostatic tissue. *Actas Urol Esp* 15: 344-350.
- 43. Zaichick V, Sviridova T, Zaichick S (1997) Zinc in human prostate gland: normal, hyperplastic and cancerous. *Int Urol Nephrol* 129: 565-574.
- 44. Galván-Bobadilla AI, García-Escamilla RM, Gutiérrez-García N, Mendoza-Magaña ML, Rosiles-Martínez R (2005) Cadmium and zinc concentrations in prostate cancer and benign prostatehyperplasia. *Rev Latinoamer Patol Clin* 52: 109-117.
- 45. Yaman M, Atici D, Bakirdere S, Akdeniz I (2005) Comparison of trace metal concentrations in malignant and benign human prostate. *J Med Chem* 48: 630-634.
- 46. Kwiatek WM, Banas A, Banas K, Podgorczyk M, Dyduch G, et al. (2006) Distinguishing prostate cancer from hyperplasia. *Acta Physica Polonica* 109: 377-381.
- 47. Guntupalli JNR, Padala S, Gummuluri AVR, Mukkineni RK, Byreddy SR, et al. (2007) Trace elemental analysis of normal, benign hypertrophic and cancerous tissues of the prostate gland using the particle-induced X-ray emission technique. *Eur J Cancer Prev* 16: 108-115.
- 48. Tohno S, Kobayashi M, Shimizu H, Tohno Y, Suwannahoy P, et al. (2009) Age-related changes of the concentrations of select elements in the prostates of Japanese. *Biol Trace Elem Res* 127: 211-227.
- 49. Kiziler AR, Aydemir B, Guzel S, Alici B, Ataus S, et al. (2010) May the level and ratio changes of trace elements be utilized in identification of disease progression and grade in prostatic cancer? *Trace Elements and Electrolytes* 27: 65-72.
- 50. Zaichick S, Zaichick V (2010) Method and portable facility for energy-dispersive X-ray fluorescent analysis of zinc content in needle-biopsy specimens of prostate. *X-Ray Spectrom* 39: 83-89.
- 51. Zaichick S, Zaichick V (2012) Trace elements of normal, benign hypertrophic and cancerous tissues of the human prostate gland investigated by neutron activation analysis. *Appl Radiat Isot* 70: 81-87.
- 52. Zaichick V, Zaichick S (2013) The effect of age on Br, Ca, Cl, K, Mg, Mn, and Na mass fraction in pediatric and young adult prostate glands investigated by neutron activation analysis. *Appl Radiat Isot* 82: 145-151.
- 53. Zaichick V, Zaichick S (2013) NAA-SLR and ICP-AES Application in the assessment of mass fraction of 19 chemical elements in pediatric and young adult prostate glands. *Biol Trace Elem Res* 156: 357-366.
- 54. Leitão RG, Palumbo A, Souza PAVR, Pereira GR, Canellas CGL, et al. (2014) Elemental concentration analysis in prostate tissues using total reflection X-ray fluorescence. *Radiation Physics and Chemistry* 95: 62-64.
- 55. Zaichick S, Zaichick V (2014) EDXRF determination of trace element contents in benign prostatic hypertrophic tissue. In: *Fundamental Interactions and Neutrons, Neutron Spectroscopy, Nuclear Structure, Ultracold Neutrons, Related Topics*. Joint Institute for Nuclear Research, Dubna (Russia): 311-316.
- 56. Denoyer D, Clatworthy SAS, Masaldan S, Meggyes PM, Cater MA (2015) Heterogeneous Copper Concentrations in Cancerous Human Prostate Tissues. *Prostate* 75: 1510-1517.
- 57. Zaichick S, Zaichick V (2015) Prostatic Tissue Level of some Androgen Dependent and Independent Trace Elements in Patients with Benign Prostatic Hyperplasia. *Androl Gynecol: Curr Res* 3:3.
- 58. Singh BP, Dwivedi S, Dhakad U, Murthy RC, Choubey VK, et al. (2016) Status and Interrelationship of Zinc, Copper, Iron, Calcium and Selenium in Prostate Cancer. *Indian J Clin Biochem* 31: 50-56.
- 59. Zaichick V, Zaichick S (2016) Trace element contents in adenocarcinoma of human prostate investigated by energy dispersive X-ray fluorescent analysis. *Journal of Adenocarcinoma* 1: 1-7.
- 60. Zaichick V, Zaichick S (2016) The Bromine, Calcium, Potassium, Magnesium, Manganese, and Sodium Contents in Adenocarcinoma of Human Prostate Gland. *J Hematology and Oncology Research* 2: 1-12.
- 61. Zaichick V, Zaichick S (2016) Trace element contents in adenocarcinoma of the human prostate gland investigated by neutron activation analysis. *Cancer Research & Oncology* 1: 1-10.
- 62. Zaichick V, Zaichick S (2016) Prostatic tissue levels of 43 trace elements in patients with prostate adenocarcinoma. *Cancer and Clinical Oncology* 5: 79-94.
- 63. Zaichick V, Zaichick S (2016) Prostatic tissue level of some major and trace elements in patients with BPH. *Jacobs Journal of Nephrology and Urology* 3: 025.
- 64. Zaichick V, Zaichick S (2016) Levels of 43 Trace Elements in Hyperplastic Prostate Tissues. *British Journal of Medicine and Medical Research* 15: 1-12.
- 65. Zaichick V, Zaichick S (2016) Chemical elemental content / Calcium ratios in tissues of human hyperplastic prostate gland. *Journal of Applied Life Sciences International* 4: 1-11.
- 66. Zaichick V, Zaichick S (2016) Distinguishing malignant from benign prostate using Br, Ca, K, Mg, Mn, and Na content in prostatic tissue. *Integrative Molecular Medicine* 3: 733-738.
- 67. Zaichick V, Zaichick S (2016) Distinguishing malignant from benign prostate using content of 17 chemical elements in prostatic tissue. *Integr Cancer Sci Therap* 3: 579-587.
- 68. Zaichick V (1997) Sampling, sample storage and preparation of biomaterials for INAA in clinical medicine, occupational and environmental

health. In: *Harmonization of Health-Related Environmental Measurements Using Nuclear and Isotopic Techniques*. IAEA, Vienna: 123-133.

- 69. Zaichick V, Zaichick S (1996) Instrumental effect on the contamination of biomedical samples in the course of sampling. *The Journal of Analytical Chemistry* 51: 1200-1205.
- 70. Zaichick V, Zaichick S (1997) A search for losses of chemical elements during freeze-drying of biological materials. *J RadioanalNuclChem* 218: 249-253.
- 71. Zaichick V (2004) Losses of chemical elements in biological samples under the dry aching process. *Trace Elements in Medicine* 5: 17-22.
- 72. Zaichick V (1995) Application of synthetic reference materials in the Medical Radiological Research Centre. *Fresenius J Anal Chem* 352: 219-223.
- 73. Korelo AM, Zaichick V (1993) Software to optimize the multi element INAA of medical and environmental samples. In: *Activation Analysis in Environment Protection*. Joint Institute for Nuclear Research, Dubna, Russia: 326-332.
- 74. Catalona WJ (1996) Clinical utility of measurements of free and total prostate-specific antigen (PSA): A review. *Prostate* 7: 64-69.
- 75. Hjertholm P, Fenger-Gron M, Vestergaard M, Christensen MB, Borre M, et al. (2015) Variation in general practice prostate-specific antigen testing and prostate cancer outcomes: An ecological study. *Int J Cancer* 136: 435-442.
- 76. Genes VS (1967) Simple methods for cybernetic data treatment of diagnostic and physiological studies. Nauka, Moscow.