

Research Article

Isolation, Anti- Biotic Sensitivity and Biochemical Characterization of Fruit Spoiling Bacteria from Different Markets of Guwahati City

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Abstract

There is potentiality for a wide range of vegetables and fruit products to become contaminated with microorganisms. The range of microorganisms associated with outbreaks linked to fresh product encompasses bacteria, viruses and parasites. This study was undertaken to investigate microbial safety aspects of some fruits such as Mango (*Mangifera indica*), Plum (*Prunus* sp.), Guava (*Psidium guajava*), Litchi (*Litchi chinensis*), Grapes (*Vitis vinifera*), Apple (*Malus domestica*) and Banana (*Musa acuminate*).

The presence of different bacteria which were responsible for spoiling fruits was confirmed by various biochemical and microbial tests. The microorganisms isolated were characterized by VITEK analysis and identified. The results showed the presence of the bacteria like *Enterobacter cloacae* complex, *Providencia rettgeri*, *Pantoea* spp and *Serratia plymuthica*. The isolated strains were then tested against 19 different antibiotics to look into their antibiotic susceptibility. The fruits in Guwahati region are mainly spoiled by *Enterobacter cloacae* complex, though other organisms are also there such as *Pantoea* spp, *Providencia rettgeri*, *Serratia plymuthica*. Methods to control these microbes need to be framed and applied for long term storage of the fruits.

Keywords: Antibiotic; Fruit Spoilage Microbes; Guwahati; Vitek

Introduction

Fruits are vital sources of nutrients to human beings. They give the body the necessary vitamin and minerals in the right proportion for human growth and development. Fruits are seasonal and highly perishable in nature. The fruits that are cultivated in a natural environment are exposed to microbial contamination. They begin to deteriorate shortly after harvest. The fruits that are cultivated in the nature are more susceptible to the microbial attacks. Fruits may become contaminated with harmful pathogens and spoilage microorganisms either during their growing in fields, orchards, vineyards, or greenhouses, or during harvesting, postharvest handling, and distribution.

Sometimes spoilage of fruits also depends on microbial growth depends on various factors such as temperature, pH and

moisture of the particular region. Biochemical and microbial spoilage are main two causes for their spoilage. Due to the spoilage, the nutritional value and the market value of the fruits also gets deteriorated. It is estimated one-fourth of the harvested fruits are spoiled before consumption. Biochemical spoilage is caused by microorganisms like bacterial, yeast and moulds. Many microorganisms that are responsible for the spoilage of fruits are also harmful to human when consumed.

These may include respiratory failure, urinary infection, food poisoning, stomachache, fever, nausea etc. Over the years, there has been an increased demand to isolate and identify the microorganisms associated with the spoilage so that it can be controlled [1,2]. This study was undertaken to isolate and identify microorganisms that are associated or responsible for the spoilage of ripened fruits sold in Narengi and Paltan bazaar markets within Guwahati region. In addition, the study investigated the susceptibility of the microorganisms to some antibiotics. *Enterobacter* species are known to be associated with high resistance to most of the

antimicrobial agents [3]. Numerous studies on the mutation and its association with drug resistance have been reported [4-7]. Similar experience in terms of resistance was also been experienced in the species isolated from food stuff [8-11]. The experiment was carried to estimate and identify the fruit spoiling microbes in Guwahati.

Materials and Methods

The experiments were conducted in the Department of Biotechnology, Assam Down Town University, Panikhaiti, Guwahati, Assam and apart with identification were carried out in Assam Down Town Hospital, India. The fruits selected for the study were: Apple (*Malus domestica*), Mango (*Mangifera indica*), Banana (*Musa acuminate*), Litchi (*Litchi chinensis*), Guava (*Psidium guava*), Grapes (*Vitis vinifera*), Plum (*Prunus cerasifera*) which are most frequently consumed by the people of the region.

Collection of Deteriorated Samples

All samples of fruits were collected from two markets of Guwahati region of, Narengi and Paltan Bazaar. The fruits selected were slightly spoiled ones. The samples were taken to the laboratory, washed and drained of water. A sterile blade and forceps were used to cut small section of the tissue containing both the healthy and the rotten portion. The cut portions were pounded with pistil and mortar to make paste. The pastes were kept in sterile tubes and labeled for future use. A total of 9 representative samples were tested.

Isolation of Bacteria

The pour-plate method according to Harigan and McCane (1990) [12] was adopted. Using standard Microbiological technique (serial dilution), a tenfold dilution of 1g of the sample was carried out in 9ml of sterile water (this was the aliquot). The exponential dilution was carried out to the fourth factor (10⁻⁴). 1ml of the fourth factor was aseptically transferred and plated in duplicate sets using sterile molten lukewarm nutrient agar which was amended with 1%. The plates were allowed to set and were incubated at 37°C for 24hours. Individual colonies were chosen and the top of an individual colony each was touched lightly with a sterile loop or needle and streak on a fresh agar plate [13,14]. The various colonies observed in the plates were distinguished on the basis of their cultural characteristics such as colony size, shape, color, consistency and hemolytic characteristics. Bacterial growth was sub-cultured on NA and was preserved.

Tests for Characterization of Bacterial Colonies

The tests to characterize the bacterial colonies were Gram staining, Catalase test, Methyl red test, Indole test, citrate utilization test, fermentation of carbohydrate test, triple sugar iron test, cugolase test, temperature assays, antibiotic tests and Vitek Test. The standard procedures for its test were performed [15-18]. As

the current study was on representative samples hence statistical analysis was not undertaken.

Results

Collection of samples (spoiled fruits) and isolation of different bacteria from spoiled fruits could be seen from (Figure 1).

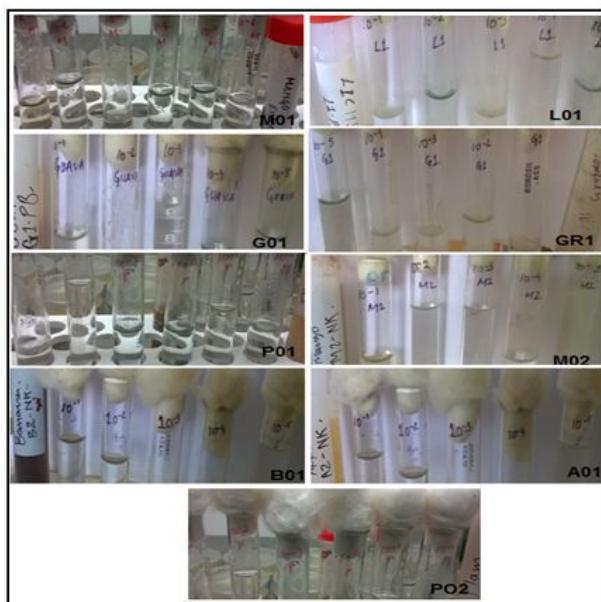


Figure 1: Serial Dilution Method.

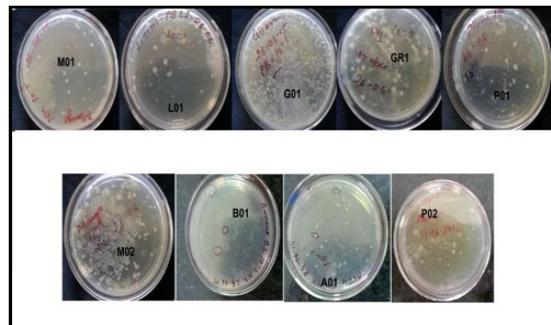


Figure 2: Pour plate method.

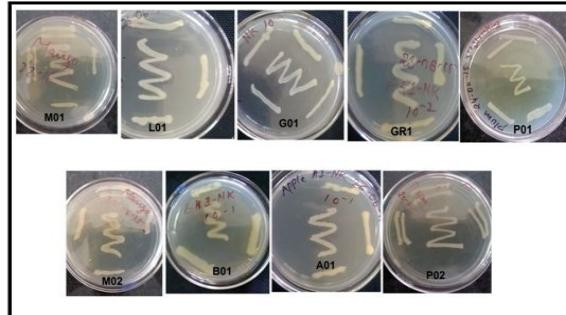


Figure 3: Streaking Method.

The results for the biochemical tests are summarized in (Table 1). Most of the bacterial strains were positive for Catalase, citrate and coagulase tests [19]. The details of remaining tests have been shown in (Table 1).

Sl. No.	Bacterial Strain	BIOCHEMICAL TEST RESULT							
		Catalase	METHYL RED TEST	INDOLE TEST	CITRATE UTILIZATION TEST	FERMENTATION OF CARBOHYDRATE	TRIPLE SUGAR IRON TEST		COAGULASE
							H ₂ S	Gas Formation	
1.	M01	P	P	N	P	P	N	P	P
2.	L01	P	P	N	P	N	N	N	P
3.	G01	P	N	P	P	N	N	P	P
4.	GR1	P	P	N	P	N	N	P	P
5.	P01	P	P	N	P	N	N	N	P
6.	M02	P	N	P	P	N	N	P	P
7.	B01	P	P	N	P	P	N	P	P
8.	A01	P	P	P	P	P	N	P	P
9.	P02	P	P	N	P	P	N	P	P

Table 1: Biochemical Tests Results.

Temperature Sensitivity Assay of Bacteria Strains

The results for temperature sensitivity shows response at 45°C for two out of the 10 studied strains of bacteria (Table 2 and Figure 4). The tests were found to be positive for all tested bacterial species at 30 and 40°C. From temperature sensitivity assay [21,22] it was found that all isolated bacterial strains grow optimally at 40°C.

Sl. No.	Bacterial strains	AREA	Temperature (°C)		
			30	40	45
1	M01	PALTAN BA-ZZAR	Y	Y	Y
2	L01		Y	Y	N
3	G01		Y	Y	N
4	GR1		Y	Y	N
5	P01		Y	Y	Y
6	M02	NARENGI	Y	Y	Y
7	B01		Y	Y	Y
8	A01		Y	Y	Y
9	P02		Y	Y	Y

Table 2: Temperature sensitivity assay of bacteria strains.

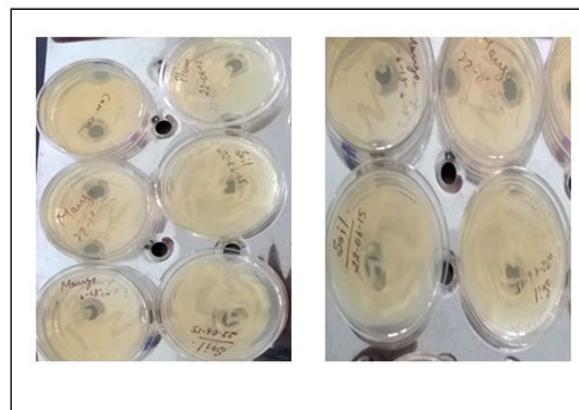


Figure 4: Temperature Sensitivity Assay.

Identification of Isolated Bacterial Strains by Vitek Technique

In terms concentration, the highest number of bacterial load was observed of *Enterobacter* species (Table 3 and Figure 5). From the Vitek Test the species were identified as six species were identified as *Enterobacter cloacae* complex [23,25], one each of *Pantoea* spp, *Providencia rettgeri*, *Serratia plymuthica*.

SL. NO	FRUITS	Bacterial Strain	PALTAN BAZZAR	Species name	Probability
1.	MANGO 1	M01		<i>Pantoea spp</i>	94%
2.	LITCHI	L01		<i>Providencia rettgeri</i>	99%
3.	GUAVA	G01		<i>Enterobacter cloacae complex</i>	97%
4.	GRAPES	GR1		<i>Enterobacter cloacae complex</i>	96%
5.	PLUM 1	P01		<i>Serratia plymuthica</i>	88%
6.	MANGO 2	M02	NARNGI BAZZAR	<i>Enterobacter cloacae complex</i>	99%
7.	BANANA	B01		<i>Enterobacter cloacae complex</i>	98%
8.	APPLE	A01		<i>Enterobacter cloacae complex</i>	99%
9.	PLUM 2	P02		<i>Enterobacter cloacae complex</i>	94%

Table 3: Result of Vitek Test.

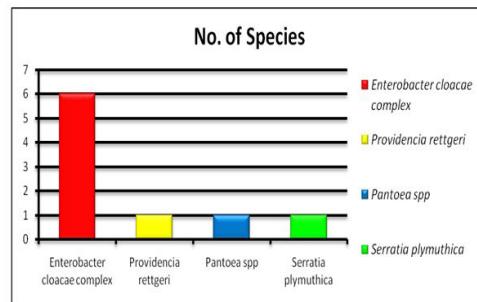


Figure 5: Graphical Representation of the No. of Species.

Antibiotic Sensitivity Assay of Bacteria Strains

The results for antibiotic tests are summarized in Table 4. From the antibiotic sensitivity test we have tested 19 antibiotics on 9 strains of bacteria that were previously isolated and seen that most of the [26] strains are resistance to some of the antibiotics.

Sl. No.	SPECIMEN NAME	VITEK TEST RESULT	ANTIBIOTIC SENSITIVITY															Alupipo 500		Monocef-O-200		Trumox CV-375		INCLAVE-375		Zefocep 200		Azomyacin 500		Metron Fort		DOXY-100		Olox-200		L-Cin 500		Moxicip		OLOX-OZ		Ceftrax-O		Ciplex-TZ		NAR-TZ		Almox 500		Cefydox-o-100 DT		Cephadex- 500		Ampicillin		Simeplomycin	
			A	M	T	Z	A1	A2	A3	A4	A5	A6	A7	A8	A9	A10	A11	A12	A13	A14	A15																																				
			1	M01	PALTAN BAZZAR	<i>Pantoea spp</i>	R	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	R	S	S	S	S	S	S	S	S	S	S																								
2	L01		<i>Providencia rettgeri</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																								
3	G01		<i>Enterobacter cloacae complex</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																								
4	GR1		<i>Enterobacter cloacae complex</i>	S	R	S	S	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																								
5	P01		<i>Serratia plymuthica</i>	S	S	S	S	S	S	-	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																								
6	M02	NARNGI BAZZAR	<i>Enterobacter cloacae complex</i>	S	S	S	S	R	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																								
7	B01		<i>Enterobacter cloacae complex</i>	S	R	S	S	S	S	S	S	S	S	S	S	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																								
8	A01		<i>Enterobacter cloacae complex</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																									
9	P02		<i>Enterobacter cloacae complex</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S																									

Table 4: Results for Antibiotic Tests.

Conclusion

Fruits provide our body with essential nutrients; vitamins etc. However, there are some bacteria that cause spoilage of fruits. A conclusion achieved was that the fruits in Guwahati region are mainly spoiled by *Enterobacter cloacae complex*, though other organisms are also there such as *Pantoea spp*, *Providencia rettgeri*, *Serratia plymuthica* [27]. Again, presence of antibiotic resistant

strains needs to be dealt carefully. The study findings are of high importance as an appropriate method for controlling these micro organism needs to be adopted as the spoilage has economical implication.

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References

1. Huis in't, Veld JH (1996) Microbial and biochemical spoilage of foods: an overview. *Int J Food Microbiol* 33: 1-18.
2. Ellis DI, Broadhurst D, Kell DB, Rowland JJ, Goodacre R (2002) Rapid and quantitative detection of the microbial spoilage of meat by fourier transform infrared spectroscopy and machine learning. *Appl Environ Microbiol* 68: 2822-2828.
3. Faheem M, Rehman MT, Danishuddin M, Khan AU (2013) Biochemical Characterization of CTX-M-15 from *Enterobacter cloacae* and Designing a Novel Non-b- Lactam-b-Lactamase. Inhibitor. *Plos One* 8: e56926.
4. Flury BB, Ellington MJ, Hopkins KL, Turton JF, Doumith M, Woodford N (2016) The differential importance of mutations within AmpD in cephalosporin resistance of *Enterobacter aerogenes* and *Enterobacter cloacae*. *Int J Antimicrob Agents* 48: 555-558.
5. Zhang D, He W, Tong Q, Zhou J, Su X (2016) Multi-omics analysis on the pathogenicity of *Enterobacter cloacae* ENHKU01 isolated from sewage outfalls along the Ningbo coastline. *Proteome Sci* 14: 15.
6. Breurec S, Bouchiat C, Sire JM, Moquet O, Bercion R, et al. (2016) High third-generation cephalosporin resistant Enterobacteriaceae prevalence rate among neonatal infections in Dakar, Senegal. *BMC Infect Dis* 16: 587.
7. Peymani A, Farivar TN, Najafipour R, Mansouri S (2016) High prevalence of plasmid-mediated quinolone resistance determinants in *Enterobacter cloacae* isolated from hospitals of the Qazvin, Alborz, and Tehran provinces, Iran. *Rev Soc Bras Med Trop* 49: 286-291.
8. Snuossi M, Trabelsi N, Ben Taleb S, Dehmeni A, Flamini G, et al. (2016) *Laurus nobilis*, *Zingiber officinale* and *Anethum graveolens* Essential Oils: Composition, Antioxidant and Antibacterial Activities against Bacteria Isolated from Fish and Shellfish. *Molecules* 21: E1414.
9. Ntuli V, Njage PM, Buys EM (2016) Characterization of *Escherichia coli* and other Enterobacteriaceae in producer-distributor bulk milk. *J Dairy Sci* 99: 9534-9549.
10. Weems MF, Dereddy NR, Arnold SR (2015) Mother's Milk as a Source of *Enterobacter cloacae* Sepsis in a Preterm Infant. *Breastfeed Med* 10: 503-504.
11. Furuhata K, Ishizaki N, Fukuyama M (2015) Antibacterial susceptibility of enterobacteriaceae isolated from raw horsemeat isolated for human consumption (basashi). *Biocontrol Sci* 20: 19-25.
12. Chueca B, Ramírez N, Arvizu-Medrano SM, García-Gonzalo D, Pagán R (2015) Inactivation of spoiling microorganisms in apple juice by a combination of essentials oils' constituents and physical treatments. *Food Sci Technol Int* 22: 389-398.
13. Choi JS, Lee YR, Ha YM, Seo HJ, Kim YH, Park SM, et al. (2014) Antibacterial effect of grapefruit seed extract (GSE) on Makgeolli-brewing microorganisms and its application in the preservation of fresh Makgeolli. *Food Sci* 79: M1159-1167.
14. Bevilacqua A, Costa C, Corbo MR, Sinigaglia M (2009) Effects of the high pressure of homogenization on some spoiling micro-organisms, representative of fruit juice microflora, inoculated in saline solution. *Lett Appl Microbiol* 48: 261-267.
15. Tyagi AK, Bukwicki D, Gottardi D, Tabanelli G, Montanari C, et al. (2014) Eucalyptua essential oil as a natural food preservative: *in vivo* and *in vitro* antieast potential. *Biomed Res Int* 2014: 969143.
16. Scaramuzza N, Berni E (2014) Heat-resistance of *Hamigeraavellanea* and *Thermoascus crustaceus* isolated from pasteurized acid products. *Int J Food Microbiol* 168-169:63-68.
17. Bevilacqua A, Speranza B, Campaniello D, Corbo MR, Sinigaglia M (2013) Inhibition of spoiling yeasts of fruits juices through citrus extracts. *J Food Prot* 76: 1753-1760.
18. Tyagi AK, Gottardi D, Malik A, Guerzoni ME (2013) Anti-yeast activity of menthe oil and vapours through *in vitro* and *in vivo* (real fruit juices) assays, *Food Chem* 137: 108-114.
19. Körpe DA, İşeri ÖD, Sahin FI, Cabi E, Haberal M (2013) High-antibacterial activity of *Urtica* spp. Seeds extracts on food and plant pathogenic bacteria, *Int J Food Sci Nutr* 64: 355-362.
20. Bevilacqua A, Corbo MR, Sinigaglia M (2010) *In vitro* evaluation of the antimicrobial activity of eugenol, limoene, and citrus extract against bacteria and yeasts, representative of the spoiling microflora of fruits juices. *J Food Prot* 73: 888-894.
21. Tournas VH, Katsoudas E (2005) Mould and yeast flora in fresh berries, grapes and Citrus fruits. *Int J Food Microbiol* 105: 11-17.
22. Silva FV, Gibbs P (2004) Target selection in designing pasteurization processes for shelf stable high-acid fruit products. *Crit Rev Food Sci Nutr* 44: 353-360.
23. Cerny G, Hennlich W, Poralla K (1984) Spoilage of fruit juice by bacilli: isolation and characterization of the spoiling microorganisms. *Z Lebensm Unters Forsch* 179: 224-227.
24. Senser F, Rehm HJ, Rautenberg E (1967) On the knowledge of fruit-juice spoiling microorganisms. II. Molds in various fruit juices. *Zentralbl Bakteriol Parasitenkd Infektionskr Hyg* 121: 736-746.
25. Ragaert P, Devlieghere F, Debevere J (2007) Role of microbiological and physiological spoilage mechanisms during storage of minimally processed vegetables; *Postharvest Biology and Technology* 44:185-194.
26. Körpe DA, İşeri ÖD, Sahin FI, Cabi E, Haberal M (2013) High-antibacterial activity of *Urtica* spp. Seeds extracts on food and plant pathogenic bacteria, *Int J Food Sci Nutr* 64: 355-362.
27. Aneja Rk, Dhiman R, Aggarwal NK, Kumar V, Kaur M (2014) Microbes Associated with Freshly Prepared Juices of Citrus and Carrots. *International Journal of Food* 2014: 1-7.