

## Research Article

# Biocontrol strategy of *Diaphania Pulverulentalis* targeting JHEH gene through Molecular cloning and Insilico analysis

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### Abstract

In insects, metamorphosis is controlled by interaction of endocrine agents mainly by Juvenile hormone and ecdysone. Juvenile Hormone Epoxide Hydrolase (JHEH), one of the major enzymes is involved in degradation of Juvenile hormone pathway and could be applied to control insects due to its irreversible reaction. The present work is focused on cloning of partial gene sequence and Insilico analysis of JHEH gene in *Diaphania pulverulentalis* (*Dp*) that is a serious insect pest on mulberry. A cDNA (800bps) encoding the partial JHEH was cloned from the mulberry leaf webber, *Diaphania pulverulentalis*. Partial *Dp*JHEH sequence contains an open reading frame encoding 244 amino acids with high degree of similarity to the reported insect JHEHs at NCBI. This is the kind of its first study on molecular cloning and sequence information of JHEH in *Diaphania pulverulentalis*.

**Keywords:** Cloning; In-silico analysis; *Diaphania pulverulentalis*; JH pathway; Juvenile hormone epoxide hydrolase

### Introduction

Juvenile Hormone (JH) is a sesquiterpenoid secreted by corpora allata in Insects. Juvenile Hormone plays an important role in physiology, development and reproduction in insect lifecycle. The JH titer in insects was regulated by JH biosynthesis and its degradation by specific enzymes [1]. JH levels are dramatically decreased at the final instar of larval growth, where the ecdysone levels are very high and responsible for pupation. The major enzymes involved in reducing and scavenging the JH before moult of last instar larval stage to pupa were Juvenile Hormone Esterase (JHE), Juvenile Hormone Epoxide Hydrolase (JHEH) and Juvenile Hormone Diol Kinase (JHDK). Formation of JH acid metabolite occurs by the hydrolysis of methyl ester of JH by JHE and hydration of epoxide moiety to JH diol is done by JHEH or JHEH acid diol will be formed by the simultaneous action of both the enzymes JHE and JHEH [2,3].

Most of the research was concentrated on the mechanism and action of JHE in the JH degradation pathway. When compared to the JHE, very little is known about JHEH though it seems to be as critical as JHE in insect development. A microsomal epoxide hydrolase known as JHEH is a hydrolytic enzyme that belongs to the family  $\alpha/\beta$  hydrolase. Due to the irreversible mechanism of JHEH on JH degradation pathway it could be applied to control insect pests [4]. JHEH genes have been characterized in many insects. In lepidopteran insect pests the JHEH genes are isolated from *Manduca sexta* [5], *Ctenocephalides felis* [6], *Heliothis virescens* [7], *Trichoplusia ni* [8], *Spilarctia obliqua* [9,10].

The insect pests attacking the silkworms and its host plant has increased and a new pest attack is being identifying regularly. These pests cause severe damage to the silkworm host plant and economic loss of Sericulture industry. Recently leaf defoliators become a serious threat to seri-farmers as they voraciously feed on young leaves of mulberry which are more important for chawki rearing. Among them *Diaphania pulverulentalis* (Hampson), commonly called as Leaf roller, has been observed as a severe pest on mulberry since 1995 in Karnataka state [11] that causes a major

damage to Mulberry tender leaves. Inhibition of JH degradation pathway causes changes in the JH titer leads to disturbances in physiology of Insects. JHEH is one of the enzymes involved in the removal of JH from the insects. In this paper, the cloning of the active sites and functional regions of JHEH sequence related with metamorphosis of last instar larvae in mulberry leaf webber - *Diaphania pulverulentalis* is reported. The obtained partial sequence of JHEH gene was analyzed by Insilico studies.

## Materials and Methods

### Insect Rearing

The larvae of *Diaphania pulverulentalis* of Mulberry were collected from infested mulberry gardens of surrounding villages of Chittoor District, Andhra Pradesh, India and rearing has done under optimum laboratory conditions. The larvae were fed with fresh V-1 variety mulberry tender leaves. They were reared in insect cage under controlled conditions (24±1°C, 75±5% RH) and a photoperiod of 16:8 [12]. The cuticles of day 2, 3 and 4 of fifth instar larvae were dissected and preceded for the RNA isolation.

### RNA isolation

The total RNA was extracted from cuticles of second, third and fourth days of fifth instar larvae of *Diaphania pulverulentalis* by TRIzol (Invitrogen) method. The tissues were weighed and frozen in liquid nitrogen. The tissue was homogenized with ceramic mortar and pestle. 1 ml of TRIzol per 100 mg of tissue was added and left for 30 min in room temperature until the sample (Tissue sample + TRIzol reagent) becomes liquefied. The samples were centrifuged at 12,000 rpm for 10 min at 4°C. Later, the supernatant was transferred into new tube and added 0.2ml of chloroform per 1 ml of TRIzol. It was mixed vigorously and centrifuged at 12,000 rpm for 15 min at 4°C. The aqueous layer was carefully separated and added 0.5 ml of Isopropanol. It was incubated in -20°C for 50 min. The precipitated RNA was collected by centrifugation at 12,000 rpm for 10 min at 4°C. The pellet was twice washed with 75% ethanol and allowed it for partial drying. The partial dried RNA pellet was dissolved in 40 µl of DEPC treated water. The quality and quantity of RNA was measured at 260/280 and 260/230 nm absorbance ratios with a spectrophotometer Nano Drop and was viewed by performing 2% Agarose gel electrophoresis (Sigma Low EEO). The RNA samples were stored at -80°C.

### First Strand cDNA synthesis

2 µg of total RNA sample was used to synthesize the single stranded cDNA. Oligo dT primer (50 µM), dNTP mixture (10 µM) and RNase free water were added to the total RNA and made up the volume to 10 µl. It was incubated for 5 min at 65°C and was immediately chilled on ice. To the above reaction mixture 5X Prime script buffer, RNase inhibitor (40U/µl), Prime Script

Reverse Transcriptase (200U/µl, Clontech) were added and made up to the volume 20µl. The reaction mixture was mixed properly by vigorous shaking and incubated it for 10 min at 30°C followed by 60 min at 42°C. Finally, the Reverse Transcriptase enzyme was deactivated by incubating at 95°C for 5 min. The sample was cooled on ice. Working condition of cDNA was checked by PCR with 18s rRNA (internal control primers). The cDNA samples were stored in -20°C.

### Primer design

The *Dp*JHEH primers were designed based on its homologous sequences from *Spodoptera litura*, *Manduca sexta*, *Heliothis virescens*, *Spilarctia obliqua*, *Helicoverpa armigera* and *Trichoplusia ni*. Primer-Blast and Primer3 tools were used to check the presence of self complementarity, hairpin formation and dimerization.

### Amplification of *Dp*JHEH sequence

A pair of degenerate primers (148F: 5' TNGAYMNTRAHGMGTGGTGGG 3', JHEH6R: 5' CCAAYWGTRTCAGGYTTSGTRGC 3') were used in the amplification of JHEH gene in *Diaphania pulverulentalis*. The PCR has been run on automated Master cycler Nexus (Applied Biosystems) using Platinum TM PCR super mix high fidelity. The volume of reaction mixture was 50 µl having 45 µl of Platinum TM PCR high fidelity mixture, 0.5 µl each of 200 µM forward primer (148F), reverse primer (JHEH6R), 1µl of cDNA and 3µl of PCR grade water. Gradient PCR was performed. The PCR conditions used were 2 min at 94°C, 40 cycles of 15 sec at 94°C, 30 sec at 46-51°C, 1min at 68°C and 2 min at 68°C.

The amplified products of expected size were analyzed on 1.5% Agarose gel (Sigma, Low EEO) in 1X Tris-Acetate-EDTA buffer (40 mM Tris, 40 mM acetic acid and 1 mM ethylene diamine tetra acetic acid) stained with Ethidium bromide at 100 V for 60 min. The obtained bands were visualized by using Gel doc system. The generated expected size bands were cut from the gel, purified and eluted by PCR cleanup/DNA gel extraction kit (TAKARA) as per manufacturer's instructions. The purified products were sequenced by Sangers dideoxy nucleotide sequencing method. The chromatogram output was checked manually for minimizing ambiguities and base calling errors.

### Insilico analysis of the generated *Dp*JHEH gene

**Translated BLAST (blastx):** The obtained nucleotide sequence similarity was determined based on BLAST search using the NCBI BLAST [13] server with comparison made to Non-redundant protein database by using blastx. The results were checked manually with respect to the percentage of identity and low e-values. The Partial sequence of *Dp*JHEH was submitted to NCBI ORF finder.

## Multiple Sequence Alignment

The nearest homologues of *Dp*JHEH which were obtained in BLASTX results, retrieved from NCBI database are subjected to MAFFT [14] to identify the conserved regions. Further the Motif finder ([www.genome.jp/tools/motif/](http://www.genome.jp/tools/motif/)) and NCBI conserved domain search [15] was used to identify the biological significance sequence patterns and conserved domains in the *Dp*JHEH ORF sequence.

## Phylogenetic Analysis

Phylogenetic relatedness of *Dp*JHEH and JHEH sequences of six lepidopteran insect pests was done by Phylogeny.fr [16] to identify the evolutionary relationship among the JHEH sequences of insect pests. The tree was generated by Neighbour Joining method using Clustal format alignment.

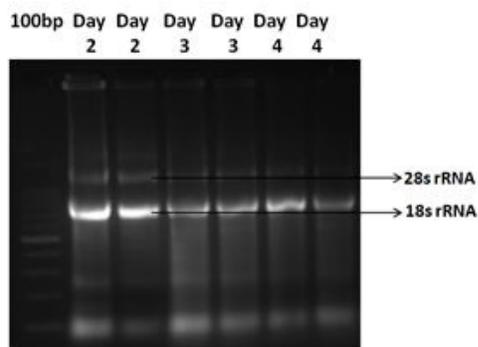
## Structural analyses

The obtained *Dp*JHEH protein sequence was subjected to PROTPARAM to explain the primary structural features like amino acid composition, Theoretical PI, Molecular weight, Instability index, grand average of hydropathicity.

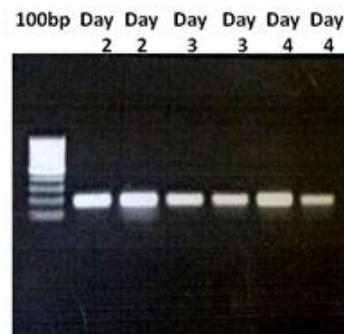
Secondary structural analysis was done by SOPMA (Self Optimized Prediction Method with Alignment) [17] to identify the percentage of alpha helices, beta turns, extended strands and random coils in *Dp*JHEH protein sequence.

## Results

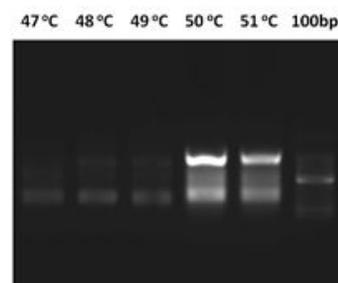
The 2% Agarose gel electrophoresis of total RNA showed high integrity bands of 28s rRNA and 18s rRNA in all the samples. The purity values of total RNA at 260/280 absorbance are within the range of 1.9 to 2.0. The gel picture of Total RNA is shown in the (Figure 1-4).



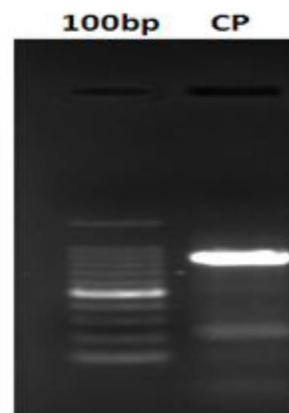
**Figure 1:** Agarose gel analysis of Total RNA samples of *Dp* fifth instar larvae.



**Figure 2:** Agarose gel analysis of PCR reaction with 18S rRNA control primers of *Dp* fifth instar larvae.



**Figure 3:** Gradient PCR (46-51°C) amplification of Partial *Dp*JHEH gene using degenerate primers 148F and JHEH6R.



**Figure 4:** Column purification (CP) of amplified PCR product of Partial *Dp*JHEH gene using degenerate primers 148F and JHEH6R.

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>D.pulverulentails_JHEH_sequence

GGCCGAAAGACTGAAAGGACGAGTGGATAAGACCGTCAAGCCCTTCAAATCAAGTTTGAAGAATCGATGGTAA

CAGACCTTCGTCAAAGACTAAAGAACCCCGATCTTTCGCGCCTCCTTGAAGGAGTGGGCTTTGAGTATGGCTT

CAATACGAAGATCATCAAAGGCTGGCTGAGCTACTGGGCCGAGAAATACAACCTCGTGAAGGGAGAAGTTCTT

GAACAAGTATCCTCAGTTCACTACTAACATTCAGGATTGGATGTGCATTTATAAGGGTACGCCCGCAGGTGTGG

GCAGGAGTTAAAGTACTTCTACTCTGGTCCACGGCTGGCCAGGTTCCGTCAGGGAGTTCTACGAGGCCATCC

CGCTTCTGACTCGCCAGACTCCAGGATACGACTTCGTGTTTGAGCTCATCGTTCCAGTCTCCCTGGTTATGTTAC

ACCGATGCTCCTGTGCGTTCGGCTTAGGCCTCCACAAACAGCGGTAATCTTCAGGAACCTGATGCACAGGCTGG

GCCACAAGAAGTTCTACGTGCAAGGAGGAGACTG GGGCGCGGGAGTCGGGGCTAGCATGGCCACCTATTCCCG

GAAGACATCTTGGGATACCACAGCAACATGATTTTTGCACAGACCAAATGCTCAACTCTCATGTACATCTTTGGGG

CTTACTCCCTCGTTCGTGTTGGAGTCAAAGCTGGCTGACCGATGTACCCTATTTCTTCATGGTTCGGGTACATA

CTTGAAGAGTTCGGCTACTTCCACCTGCAAGCCACCAAACCTG
    
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Figure 5: Amplified partial sequence (800bp) of DpJHEH gene.

The examination of cDNA working condition with 18S rRNA internal control primers was showed with well amplified bands with 200bp size. The gel picture was shown in the (Figure 2). The gradient PCR results inferred that expected amplicon size with 800 bps of *DpJHEH* was amplified at 50°C annealing temperature. The PCR gel picture and purified product of PCR was shown in the (Figure 3 & 4) respectively. The amplified partial *DpJHEH* was shown in (Figure 5).

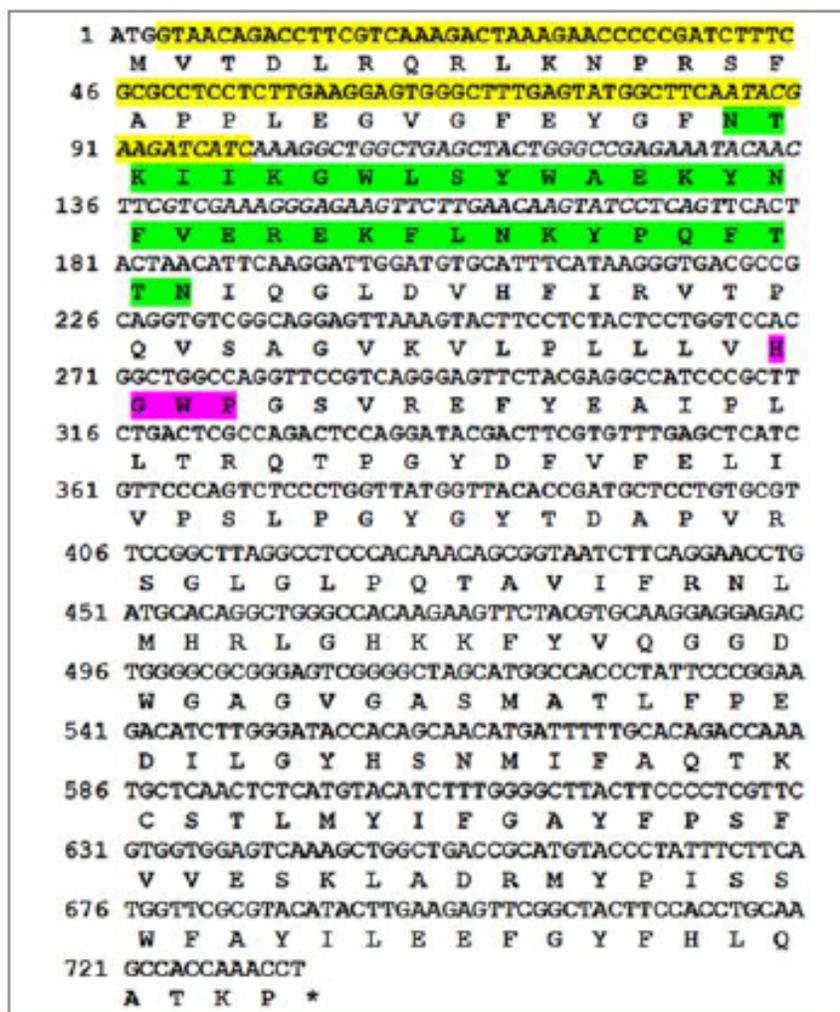
The partial *DpJHEH* showed 71% homology with JHEH sequence of *Spodoptera litura*, 70% identity with *Helicoverpa armigera*, 69% identity with *Spodoptera exigua*, 68% with *Amyelois transitella*, 67% with *Papilio xuthus* and 66% with *Heliothis virescens*. The first ten homologues of JHEH gene of *Diaphania pulverulentalis* based on blastx results were shown in the (Table 1).

BLAST hit	Max score	Total score	Query coverage	E-Value	Identity
Juvenile hormone epoxide hydrolase-like [ <i>Spodoptera litura</i> ]	417	417	99%	1e-142	71%
Juvenile hormone epoxide hydrolase-like [ <i>Helicoverpa armigera</i> ]	411	411	99%	3e-140	70%
Juvenile hormone epoxide hydrolase [ <i>Spodoptera exigua</i> ]	412	412	99%	2e-140	69%
PREDICTED: juvenile hormone epoxide hydrolase-like [ <i>Amyelois transitella</i> ]	406	406	99%	2e-138	68
Hypothetical protein B5V51_6818 [ <i>Heliothis virescens</i> ]	399	399	99%	2e-135	68

PREDICTED: juvenile hormone epoxide hydrolase-like [ <i>Papilio xuthus</i> ]	393	393	99%	3e-133	67
Microsomal epoxide hydrolase [ <i>Heliothis virescens</i> ]	393	393	99%	4e-133	66
PREDICTED: juvenile hormone epoxide hydrolase-like [ <i>Papilio polytes</i> ]	392	392	98%	1e-132	67
PREDICTED: juvenile hormone epoxide hydrolase-like isoform X2 [ <i>Papilio machaon</i> ]	391	391	99%	2e-132	67
PREDICTED: juvenile hormone epoxide hydrolase-like isoform X1 [ <i>Papilio machaon</i> ]	390	390	99%	3e-132	67

**Table 1:** Blastx results against non-redundant protein database for partial *DpJHEH* gene sequence.

The ORF sequence of partial *DpJHEH* generated by NCBI ORF Finder was showed in (Figure 6,7).



**Figure 6:** ORF sequence of *DpJHEH* [HGWP motif-Pink; EHN domain (2-99) -Yellow; AbH domain (86-175) - Italic;  $\alpha/\beta$  hydrolase domain (85-187)-Green].

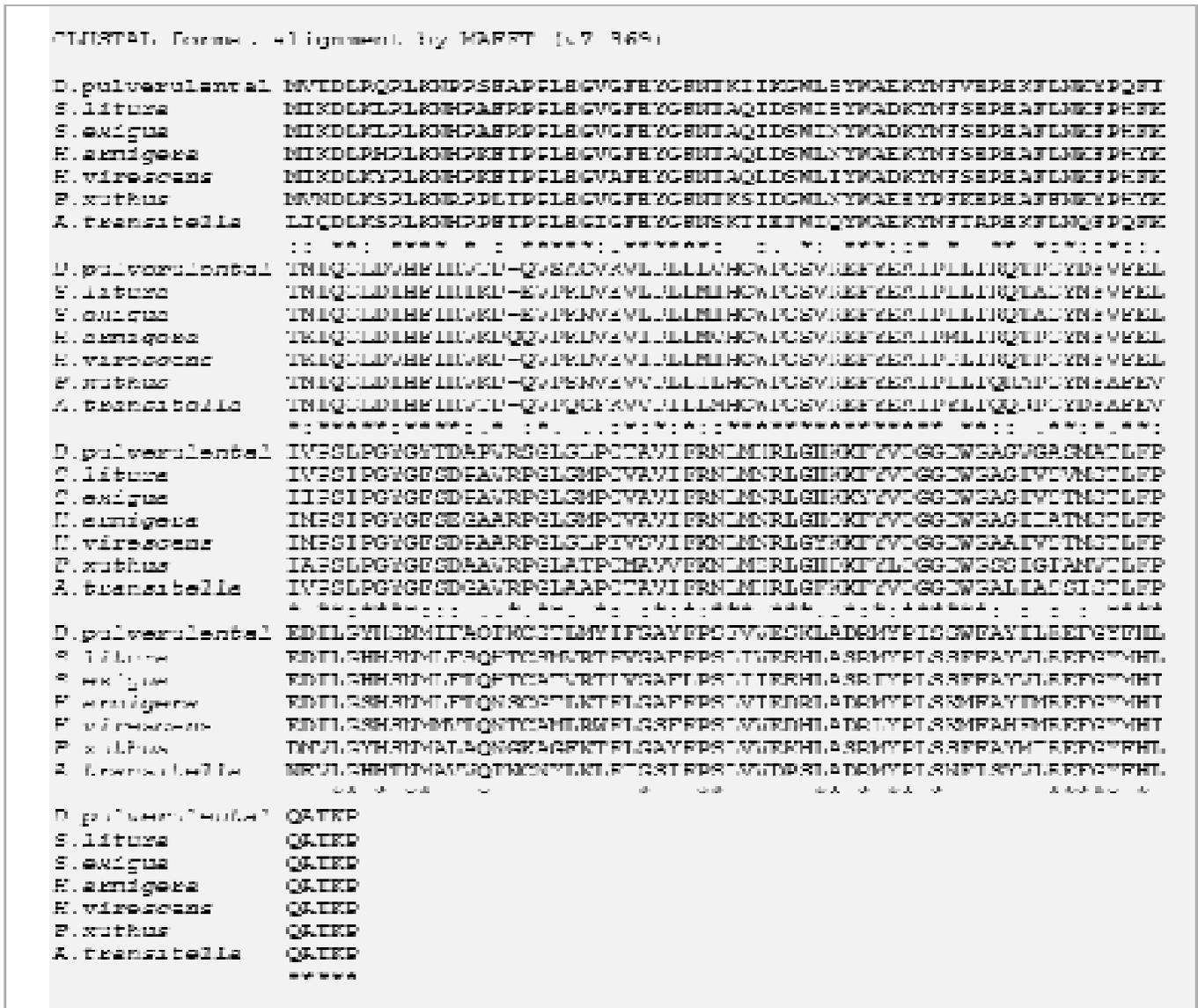
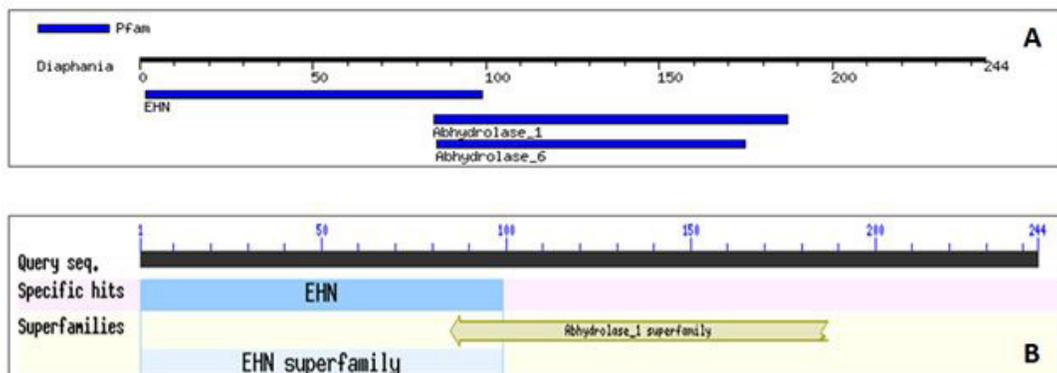


Figure 7: Multiple sequence alignment of *DpJHEH* by MAFFT (Clustal format).

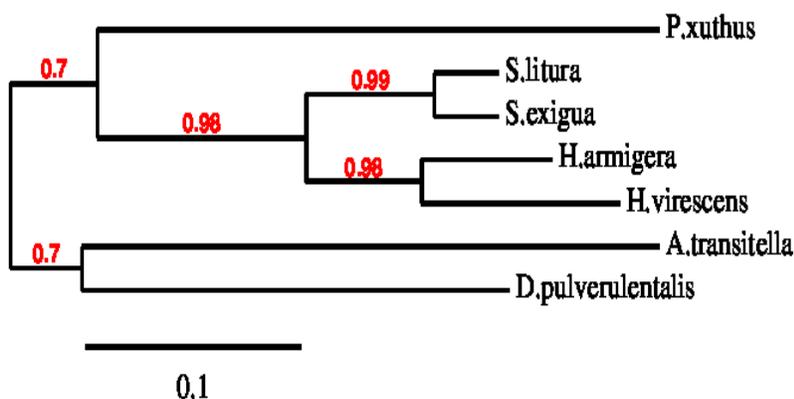
The comparison between the homologues of *DpJHEH* by MAFFT showed the HGWP motif, Epoxide hydrolase N-terminus (EHN-Position 2-99), alpha/beta hydrolase fold (Abhydrolase\_1-Position 85-187) and Alpha/beta hydrolase family (Abhydrolase\_6-Position 86-175). The multiple sequence alignment was shown in (Figure 7).

Three motifs Epoxide Hydrolase N-terminus (EHN-Position 2-99), alpha/beta hydrolase fold (Abhydrolase\_1-Position 85-187) and Alpha/beta hydrolase family (Abhydrolase\_6-Position 86-175) were identified in *DpJHEH* ORF sequence. The results of NCBI conserved domain search showed two functional domains EHN and Abhydrolase\_1 super family in translated protein sequence of *DpJHEH*. The results were showed in (Figure 8A & B).



**Figure 8:** NCBI Conserved domain search for *DpJHEH*. **A).** Three motif regions identified in Partial *DpJHEH* sequence by Motif finder. **B).** Two conserved domains regions identified in Partial *DpJHEH* by NCBI conserved Domain search.

The generated phylogenetic tree showed that the partial *DpJHEH* sequence is evolutionarily closer to the JHEH sequence of *Amyelois transitella*. The genetic divergence from the common ancestor can be observed in between JHEH sequences of *Diaphania* and *Spodoptera* species. The tree view of Phylogeny.fr was as shown in the (Figure 9).



**Figure 9:** Phylogram of *DpJHEH* sequence with other lepidopteran JHEH sequences.

The PROTPARAM output reveals that the *DpJHEH* contains 244 amino acids, 27822.19 molecular weight, 9.01 theoretical PI, 36.91 instability index, -0.007 GRAVY. The instability index of the sequence is below as 40. Henceforth the *DpJHEH* protein was predicted as stable. The protein sequence of *DpJHEH* showed 33.6% of alpha helices, 19.26% extended strands, 9.84% beta turns and 37.30% random coils. The sequence and secondary structure representation (Figure 10A), the graphical representation of secondary structure (Figure 10B) and amino acid composition (Figure 10C) of *DpJHEH* are shown below.



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